Belgian Society of Human Genetics
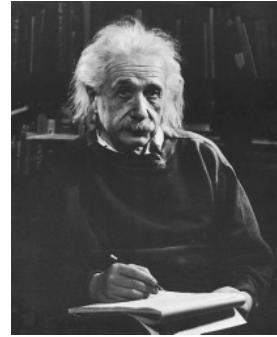
Of Mice and Human Genetics

Frank Kooy

Permanent Education Course in Human Genetics
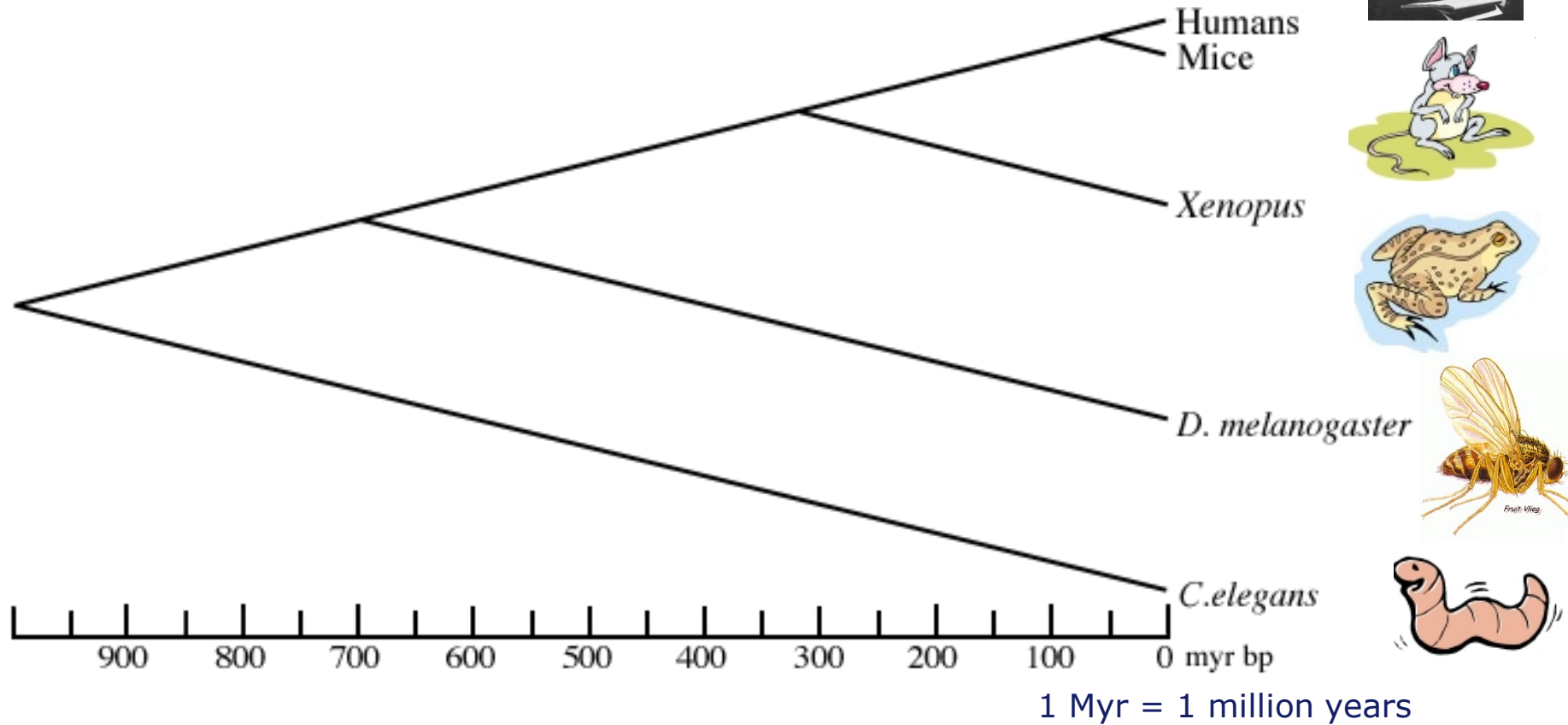
Universiteit Antwerpen

Cognitive Genetics
Centre of Medical Genetics
University of Antwerp

Why study mice

Evolutionary relationships

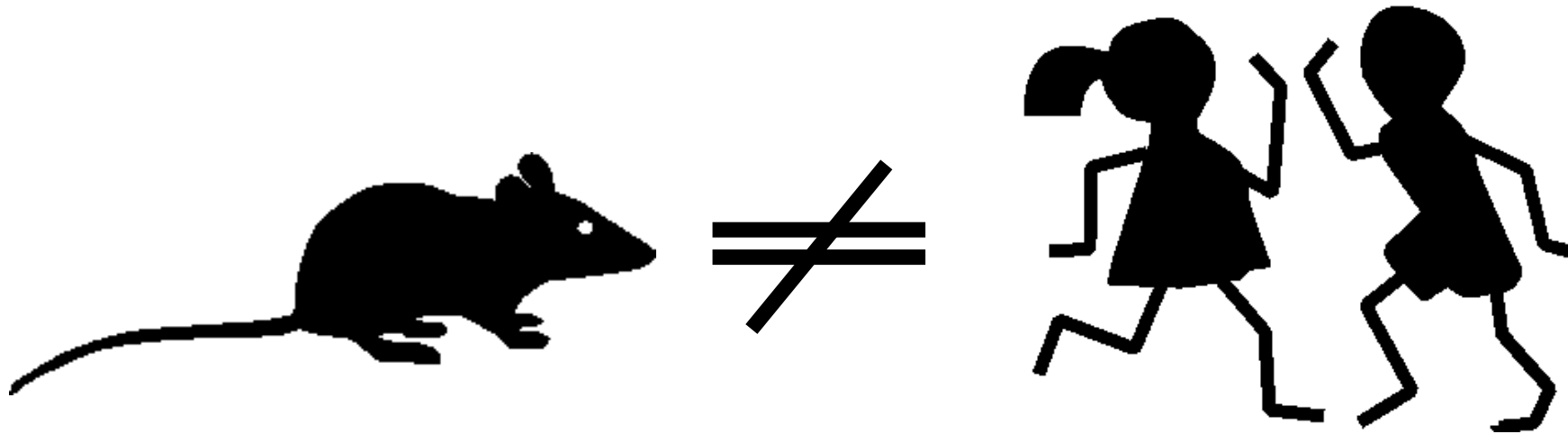900   800   700   600   500   400   300   200   100   0 myr bp

1 Myr = 1 million years

Why study mice ?

•Allow Genetic Manipulation
•Unlimited Numbers can be Bred
•Control of Genetic Background
•Availability of all Organs/Tissues
•Can be used for drug testing
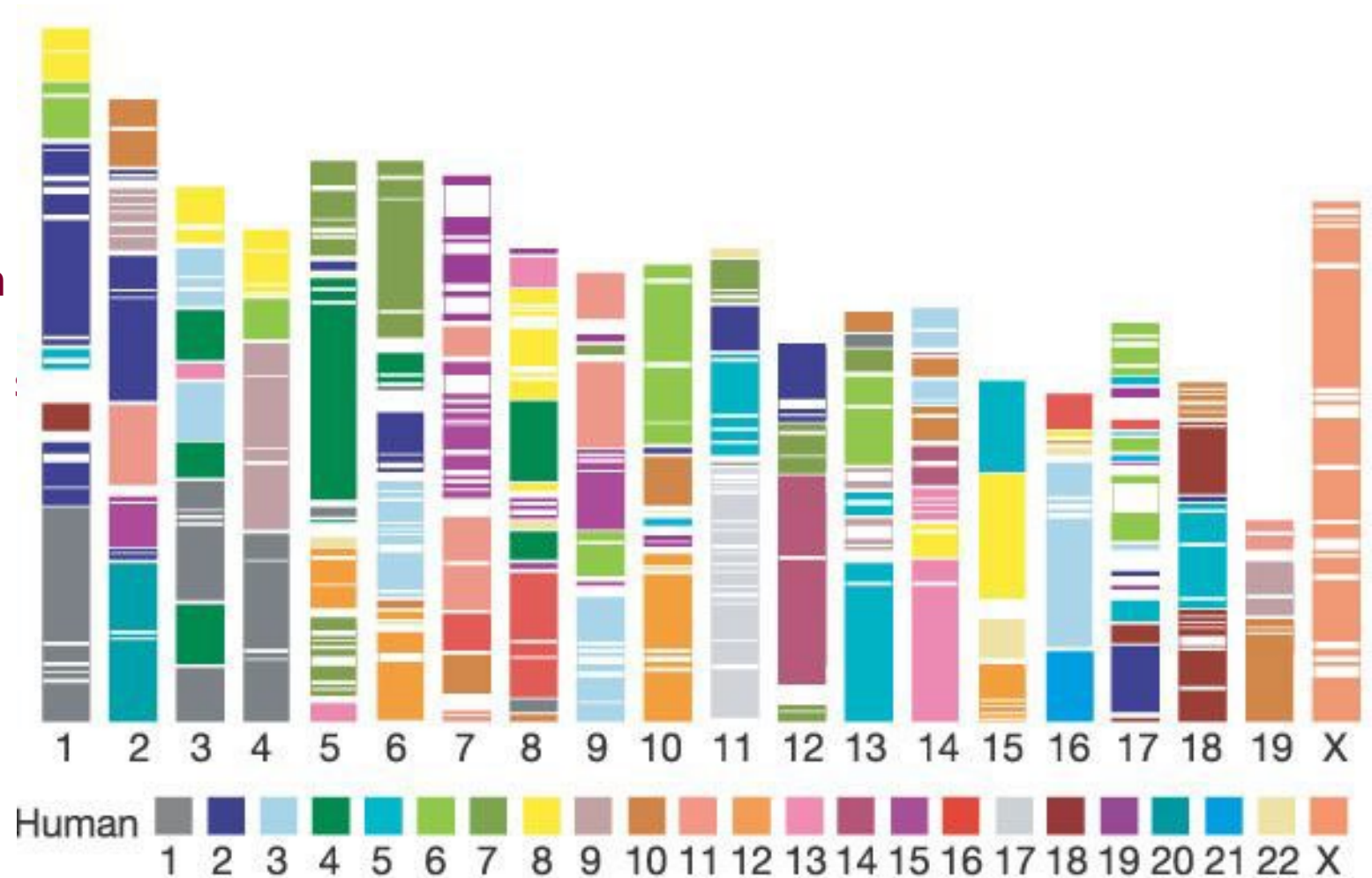
A mouse is not a man

What is in their genome?

Over 90% of the mouse and human genomes can be partitioned into corresponding regions of conserved

Only 1% of mouse genes has no identifiable orthologue in man.

Genetic modification: knockout mouse

- Knockout mouse   Inactivation of coding sequence at defined position in genome

1987-89
The first knockout mouse

Teams led by Martin Evans, Oliver Smithies and Mario Capecchi create the first 'knockout' mice, by selectively disabling a specific target gene in embryonic stem cells. The three receive the Lasker Award in 2001 for this achievement, and the technique goes on to be used to create several thousand knockout mice.

•Advantages: Specific inactivation of gene of interest

•Disadvantage: technically complicated

The Nobel Prize
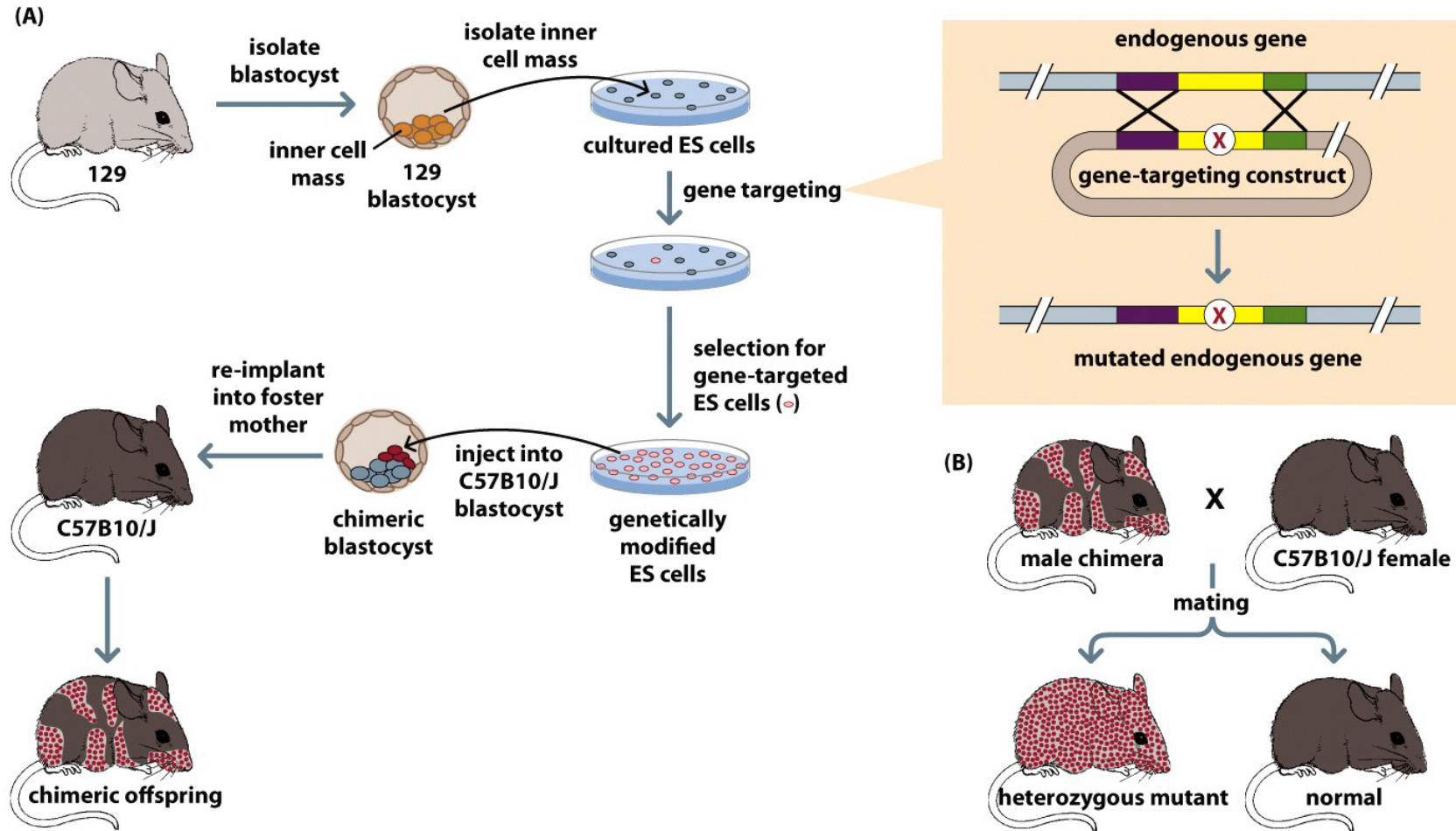in Physiology or Medicine 2007

Generation of a knockout mouse

Figure 20.6 Human Molecular Genetics, 4ed. (© Garland Science)

Generation of a knockout mouse

Fragile X mouse

Obese mouse (taconic)

gene targeting

chromosomal DNA

neo

gene-targeting construct

tk

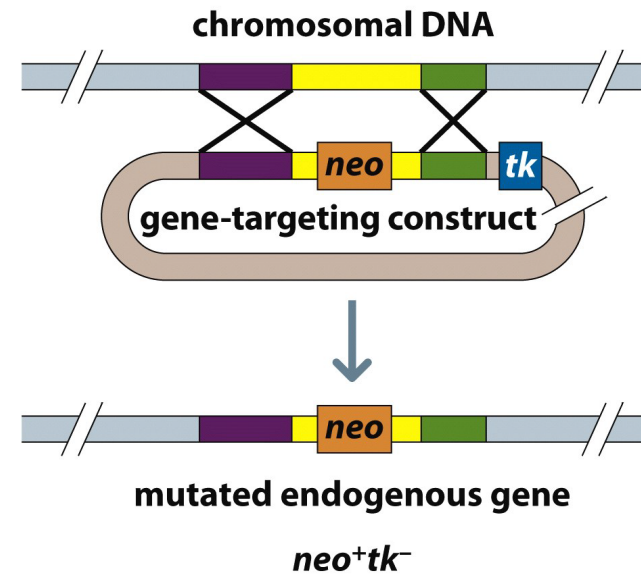mutated endogenous gene

neo⁺tk⁻

Figure 20.7a Human Molecular Genetics, 4ed. (© Garland Science)

Conditional knockout mouse

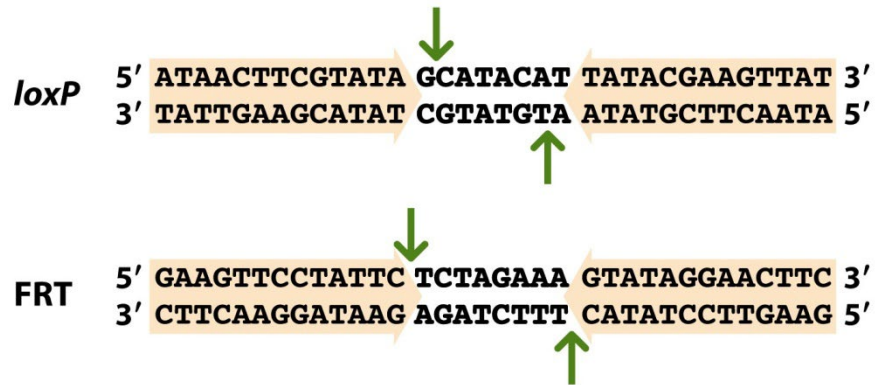Cre-recombinase and flippase are site specific recombinases

loxP
5' ATAACTTCGTATA GCATACAT TATACGAAGTTAT 3'
3' TATTGAAGCATAT CGTATGTA ATATGCTTCAATA 5'

FRT
5' GAAGTTCCTATTC TCTAGAAA GTATAGGAACTTC 3'
3' CTTCAAGGATAAG AGATCTTT CATATCCTTGAAG 5'

Figure 20.10 Human Molecular Genetics, 4ed. (© Garland Science)

mouse with target locus **A** flanked by *loxP* sites

transgenic mouse with *Cre* genes linked to a tissue-specific promoter, **P**

offspring with floxed target plus *Cre* transgene

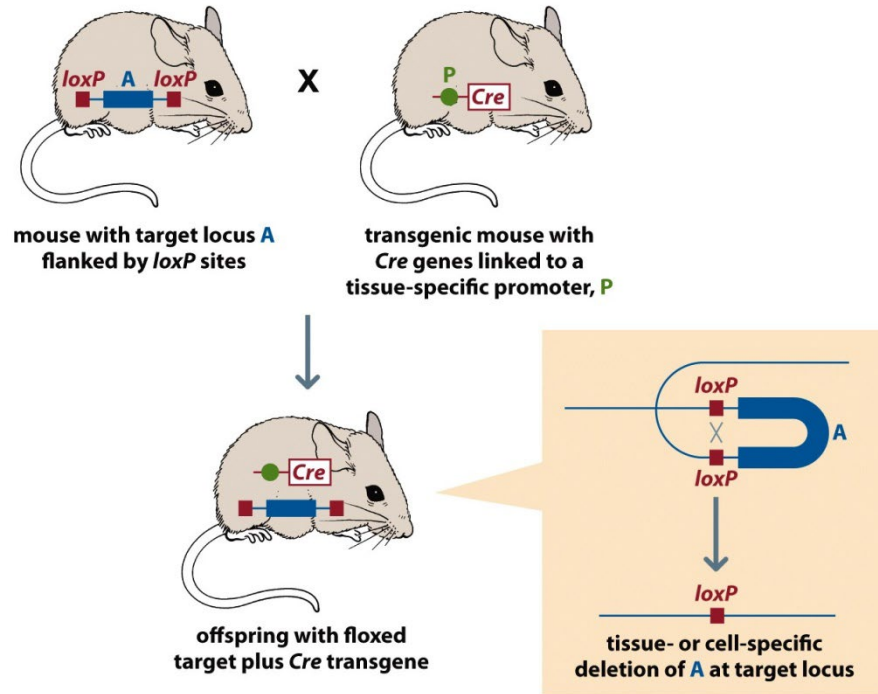tissue- or cell-specific deletion of **A** at target locus

Figure 20.11 Human Molecular Genetics, 4ed. (© Garland Science)

Transgenic mouse

- Transgenesis   Introduction of genetic material at random position in genome

•cDNA with different promotors

•Large genomic inserts with gene of interest (Cosmids, PAC, BAC, YAC) and natural promotor

•Advantage: relatively simple technology
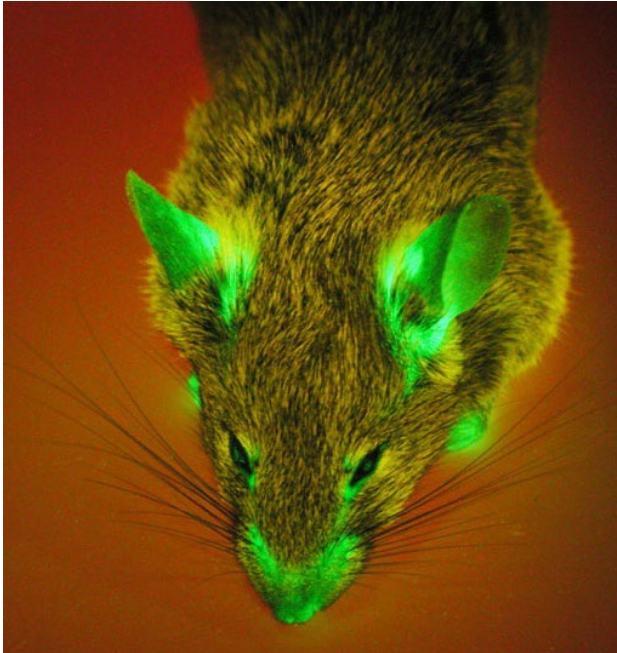
•Disadvantage: need to analyze several lines

nature

GIGANTIC MICE
INJECTED WITH GROWTH HORMONE GENES

A MILLISECOND PULSAR

1982
First transgenic mouse
A team led by Richard Palmiter and Ralph Brinster fuse elements of a gene that can be regulated by dietary zinc to a rat growth-hormone gene, and inject it into fertilized mouse embryos. The resulting mice, when fed with extra zinc, grow to be huge, and the technique paves the way for a wave of genetic analysis using transgenic mice.

Transgenesis: not limited to mice

Green fluorescent protein (GFP) transgenic

Glowfish

Generation of transgenic mice

female pronucleus male pronucleus DNA solution
microinjection pipette
holding pipette

nick in chromosomal DNA
DNA clone
integration
transgene

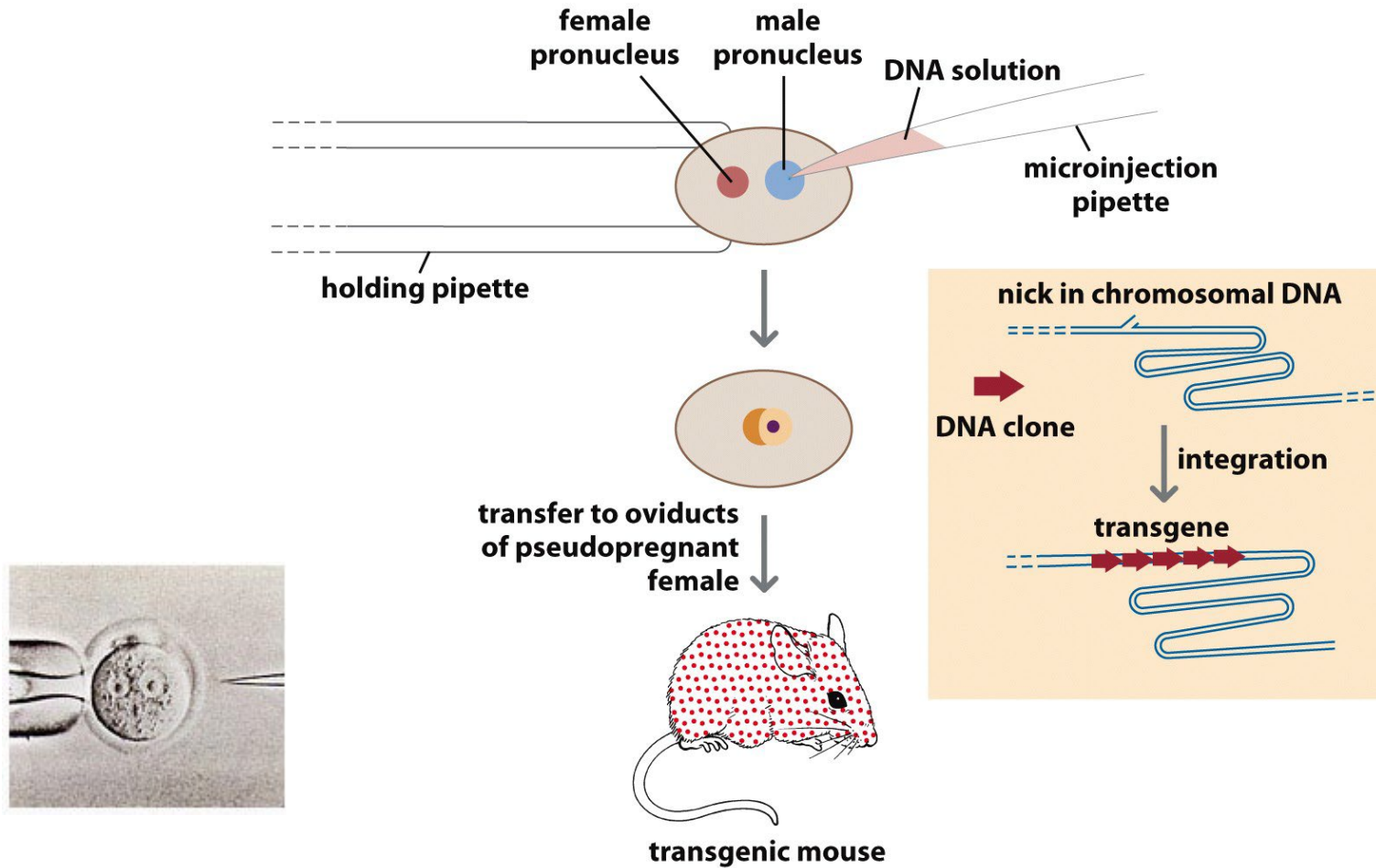transfer to oviducts of pseudopregnant female

transgenic mouse

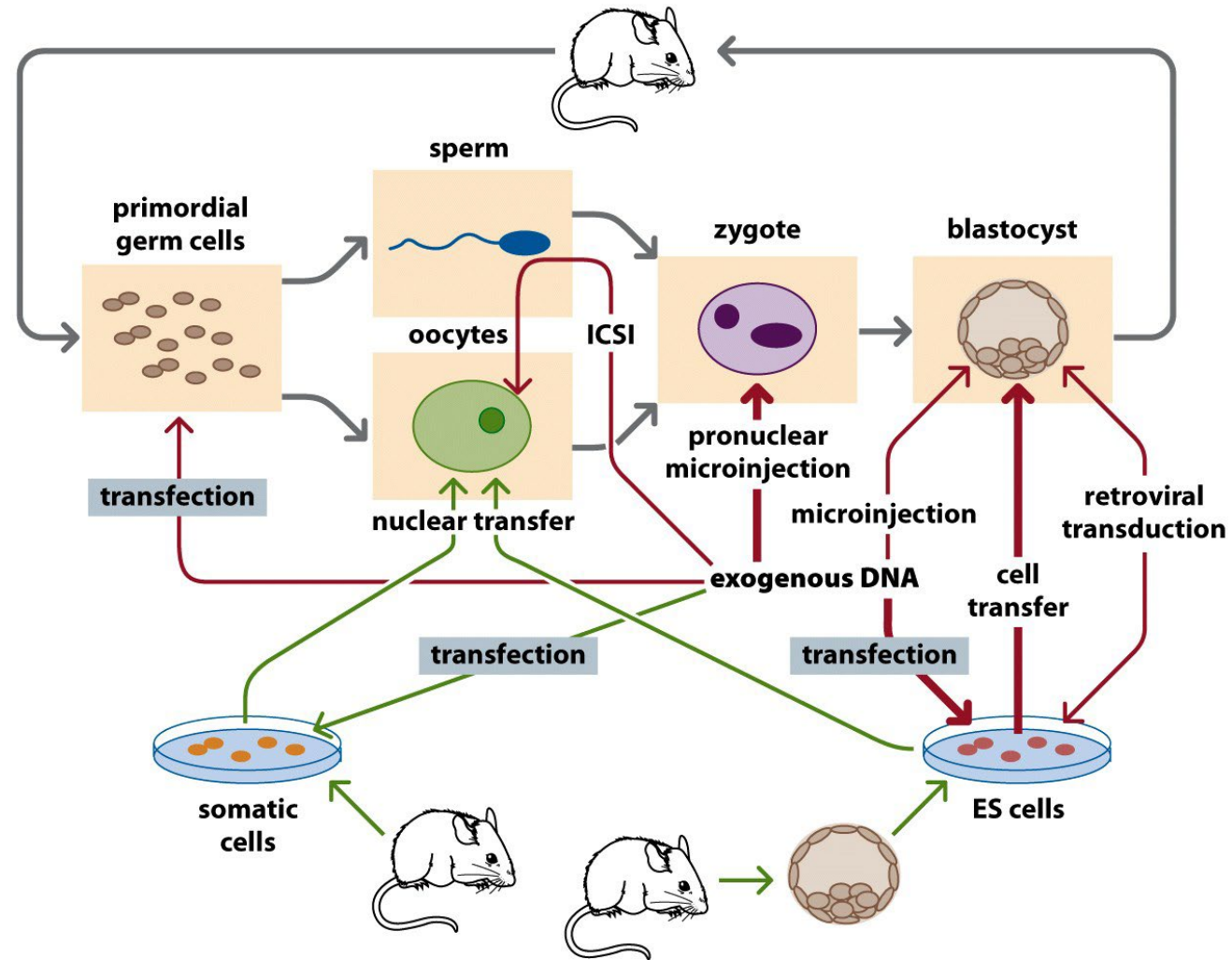Figure 20.3 Human Molecular Genetics, 4ed. (© Garland Science)

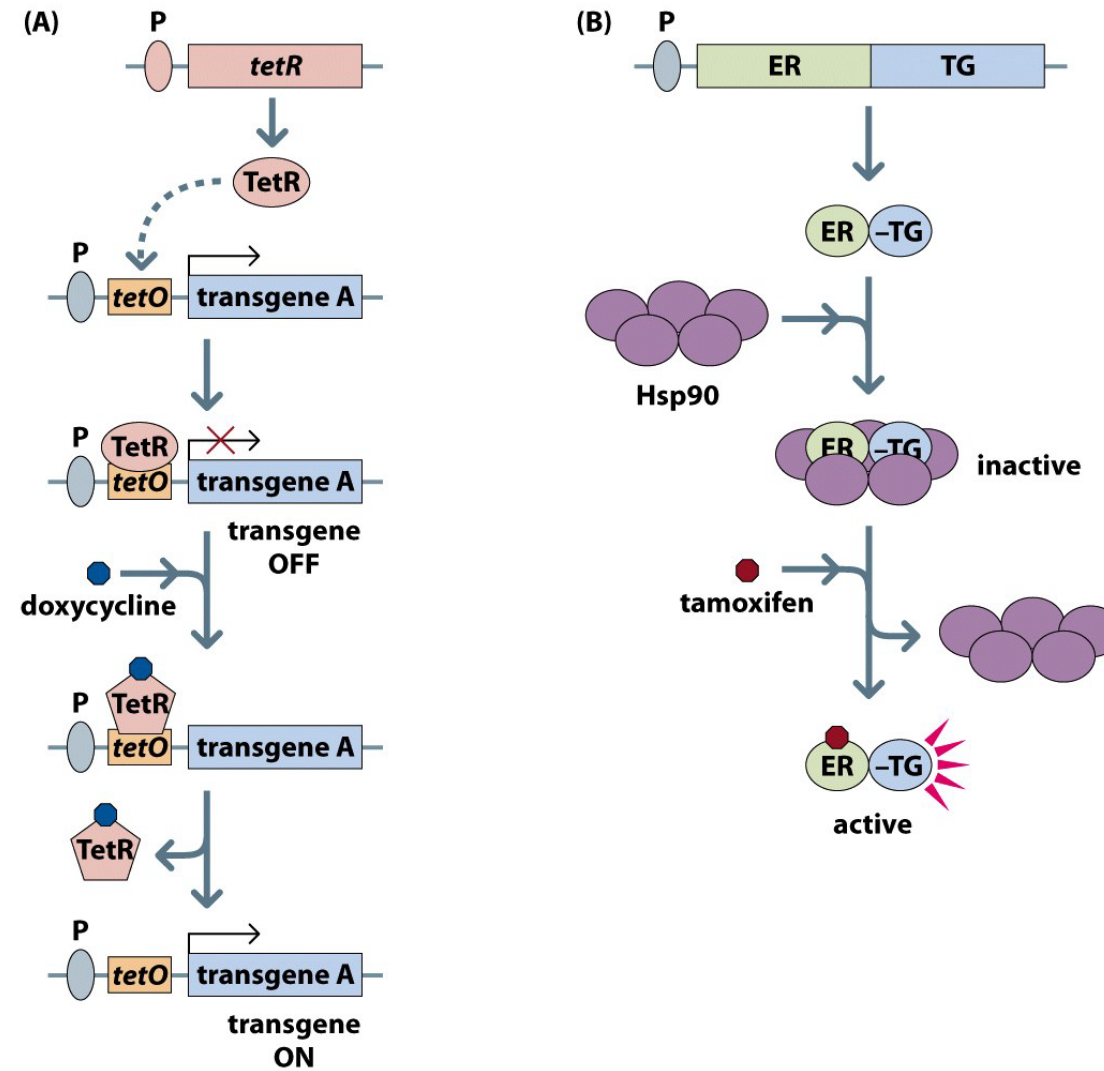Figure 20.2 Human Molecular Genetics, 4ed. (© Garland Science)

Inducible transgenic mice

Figure 20.5 Human Molecular Genetics, 4ed. (© Garland Science)

"Getting" a knockout

- Generate one
  – yourselves, in collaboration with specialized University laboratories or commercial

$$ENU \quad CH_3 - CH_2 - N - C - NH_2$$

- Seach databases (Jackson) and literature

- Seach chemically induced (ENU) mutant databases
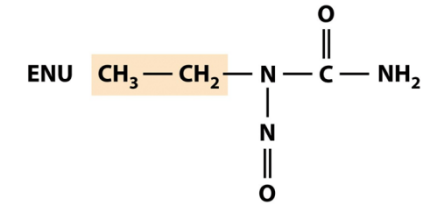
HelmholtzZentrum münchen
German Research Center for Environmental Health

Home   Imprint   Sitemap   Contact   Search   OWA   Print version

group Molecular Endocrinology
group Gene Regulation
group Functional Genetics
» start functional genetics
» ENU screen

institute of experimental genetics (ieg)

- Seach random mutagenesis databases (genetrap)

Genetrap principle

Figure 3. The conventional trapping process.

1. Random insertion of a gene trap vector into a genomic locus

Gene trap vector insertion after electroporation

2. Integration of a vector into an intron of an expressed gene

3. Transcription of the "trapped" gene results in a truncated mRNA that will translate into a selectable, tagged protein

Interruption of endogenous splicing by a polyA signal in the vector

mRNA product coding for a fusion protein

Fusion protein containing the selection or reporter protein and a fragment of the original protein.

Primers that bind within the trap cassette are used to sequence the mRNA product in the 3' to 5' direction. This sequence is used to identify the trapped gene.

primer

5' RACE sequencing

Figure 2. Schematic of a standard gene trap vector.

SA Trap cassette pA

- splice acceptor sequence
- selection/reporter construct (e.g. βgeo)
- polyadenylation signal

International Gene Trap Consortium

About IGTC

Gene trapping is a high-throughput approach that is used to introduce insertional mutations across the genome in mouse embryonic stem (ES) cells. In addition to generating standard loss-of-function alleles, newer gene trap vectors offer a variety of post-insertional modification strategies for the generation of other experimental alleles.

The International Gene Trap Consortium (IGTC) represents all publicly available gene trap cell lines, which are available on a non-collaborative basis for nominal handling fees. Researchers can search and browse the IGTC database for cell lines of interest using accession numbers or IDs, keywords, sequence data, tissue expression profiles and biological pathways.

INFORMATION

DATA ACCESS

TUTORIALS

REQUEST ES
CELL LINES
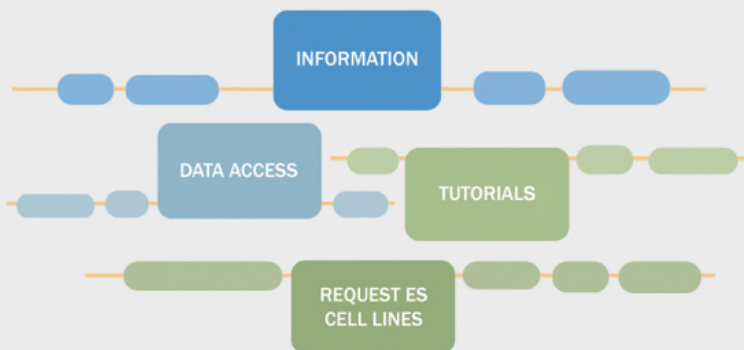
Statistics and News

› **Statistics: Mar 11, 2024**

IGTC cell lines in database:121705 cell lines
Pipeline Status:116789 (95.96%) processed cell lines
Last dbGSS deposit:2014-02-19
Ensembl version:54, NCBI m37, IGTC cell line coverage 12393 (39.33%) genes
Entrez version:NCBI m37, IGTC cell line coverage 16377 (14.91%) genes
More statistics...

› **The International Gene Trap Consortium is moving to a new home in the Mouse Biology Program at UC Davis**

Oct 14, 2010

The International Gene Trap Consortium (IGTC) web portal, www.genetrap.org, is moving to a new home at the Mouse Biology Program (MBP) at the University of California, Davis. The move became effective October 1, 2010 and was designed to enhance IGTC's longevity of services by providing a seamless transition of support to MBP's scientific and informatics staff and in-house web services infrastructure.

› **Genomic Sequence Alignment Images**

Jul 29, 2010

Individual cell line and gene pages have long included an image depicting how cDNA cell lines align to the trapped gene. As most IGTC cell lines are genomic traps, we are introducing new genomic alignment images. Now both cDNA and genomic traps are aligned against the associated gene transcripts and viewable in a scrollable Scalable Vector Graphics (SVG) image.

› **IGTC Running on New Hardware**

May 27, 2010

The IGTC web site is now running on a Red Hat Enterprise Linux server cluster.

All news and events

ARTICLE

A conditional knockout resource for the genome–wide study of mouse gene function

William C. Skarnes[1], Barry Rosen[1], Anthony P. West[1], Manousos Koutsourakis[1], Wendy Bushell[1], Vivek Iyer[1], Alejandro O. Mujica[1]†, Mark Thomas[1], Jennifer Harrow[1], Tony Cox[1], David Jackson[1], Jessica Severin[1]†, Patrick Biggs[1]†, Jun Fu[2], Michael Nefedov[3], Pieter J. de Jong[3], A. Francis Stewart[2] & Allan Bradley[1]
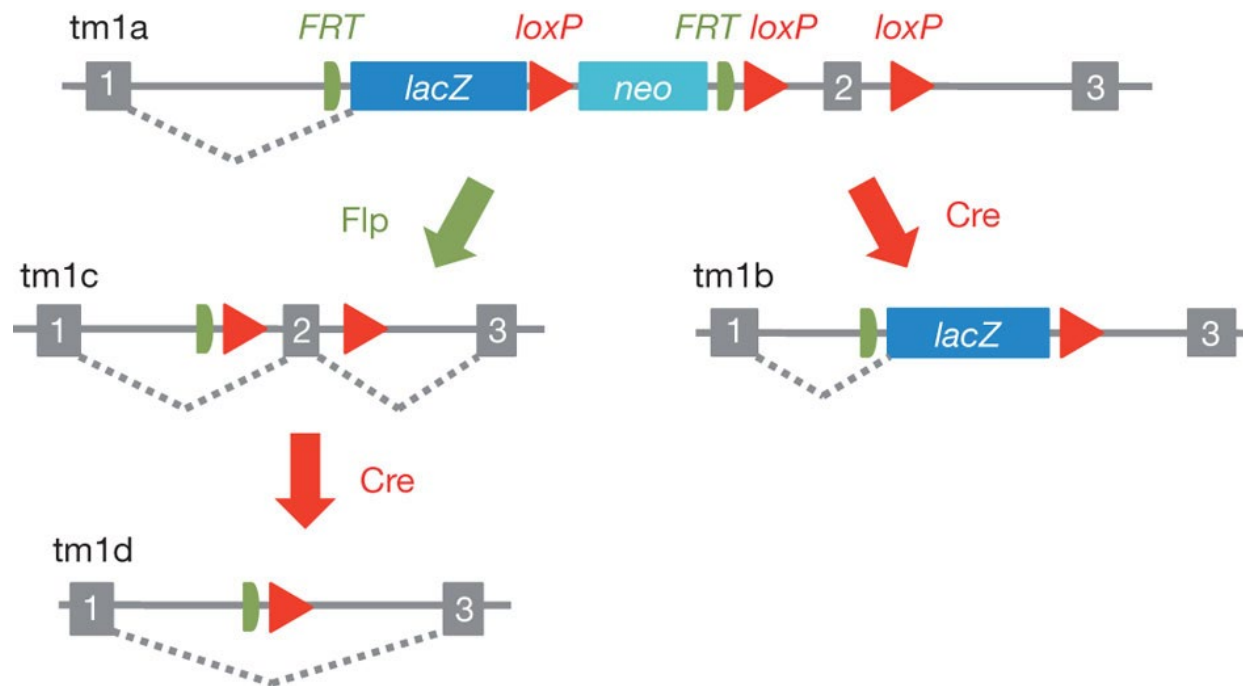
The 'knockout-first' allele (tm1a) contains an IRES:*lacZ* trapping cassette and a floxed promoter-driven *neo* cassette inserted into the intron of a gene, disrupting gene function. Flp converts the 'knockout-first' allele to a conditional allele (tm1c), restoring gene activity. Cre deletes the promoter-driven selection cassette and floxed exon of the tm1a allele to generate a *lacZ*-tagged allele (tm1b) or deletes the floxed exon of the tm1c allele to generate a frameshift mutation (tm1d), triggering nonsense mediated decay of the deleted transcript.
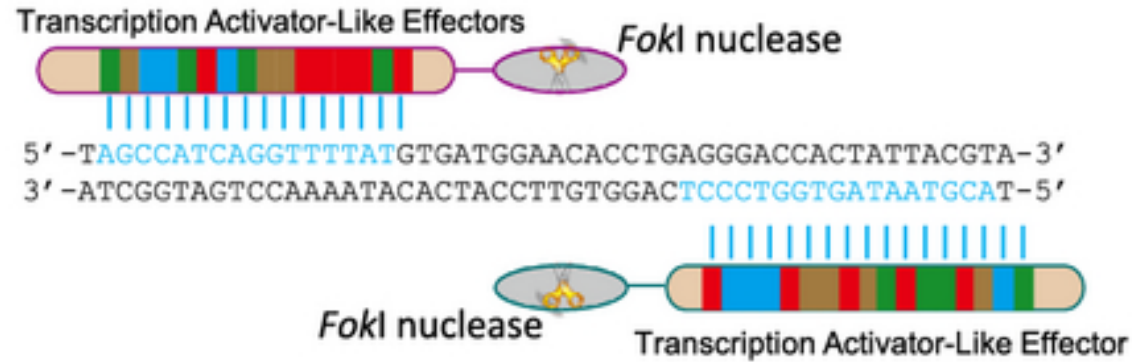
Novel methods:
Zinc-finger nucleases

Novel methods:
Transcripion Activator-Like Effector Nucleases

TAL effector nucleases (TALENs)

Transcription Activator-Like Effectors

FokI nuclease

5'-TAGCCATCAGGTTTTATGTGATGGAACACCTGAGGGACCACTATTACGTA-3'
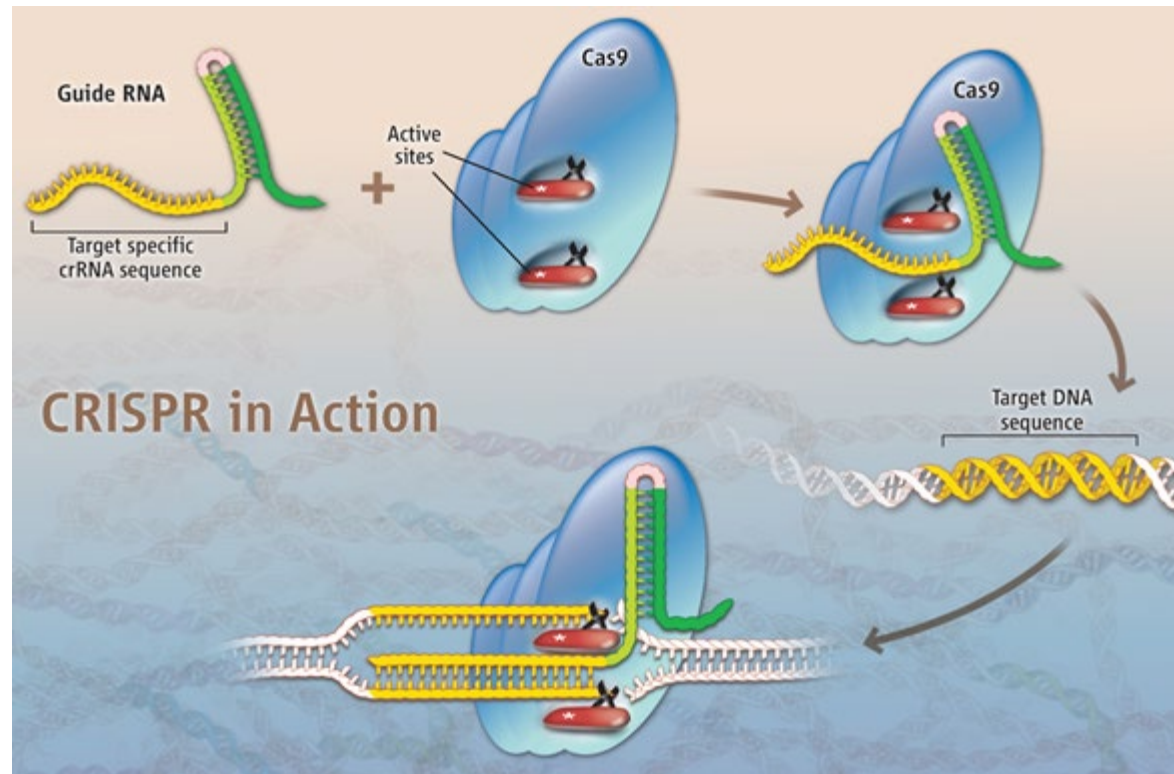3'-ATCGGTAGTCCAAAATACACTACCTTGTGGACTCCCTGGTGATAATGCAT-5'

FokI nuclease

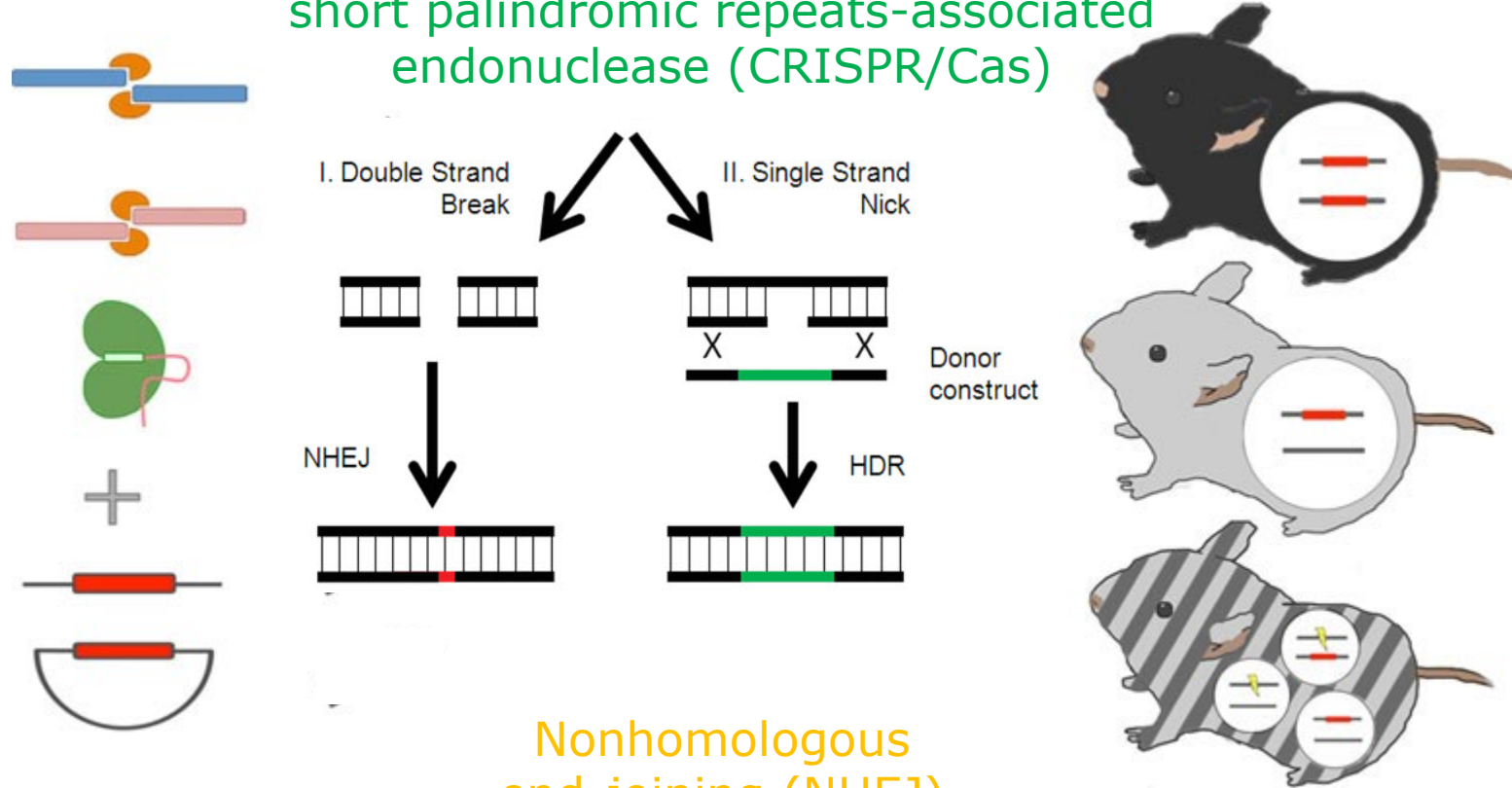Transcription Activator-Like Effector

Novel methods:
Clustered, regularly interspaced, short palindromic repeats-associated endonuclease (CRISPR/Cas)

Transcription activator-like effector nucleases (TALENs), Zinc-finger nucleases (ZFNs). Clustered, regularly interspaced, short palindromic repeats-associated endonuclease (CRISPR/Cas). Nonhomologous end-joining (NHEJ). Double-strand break (DSB) and subsequent homology-directed repair (HDR)

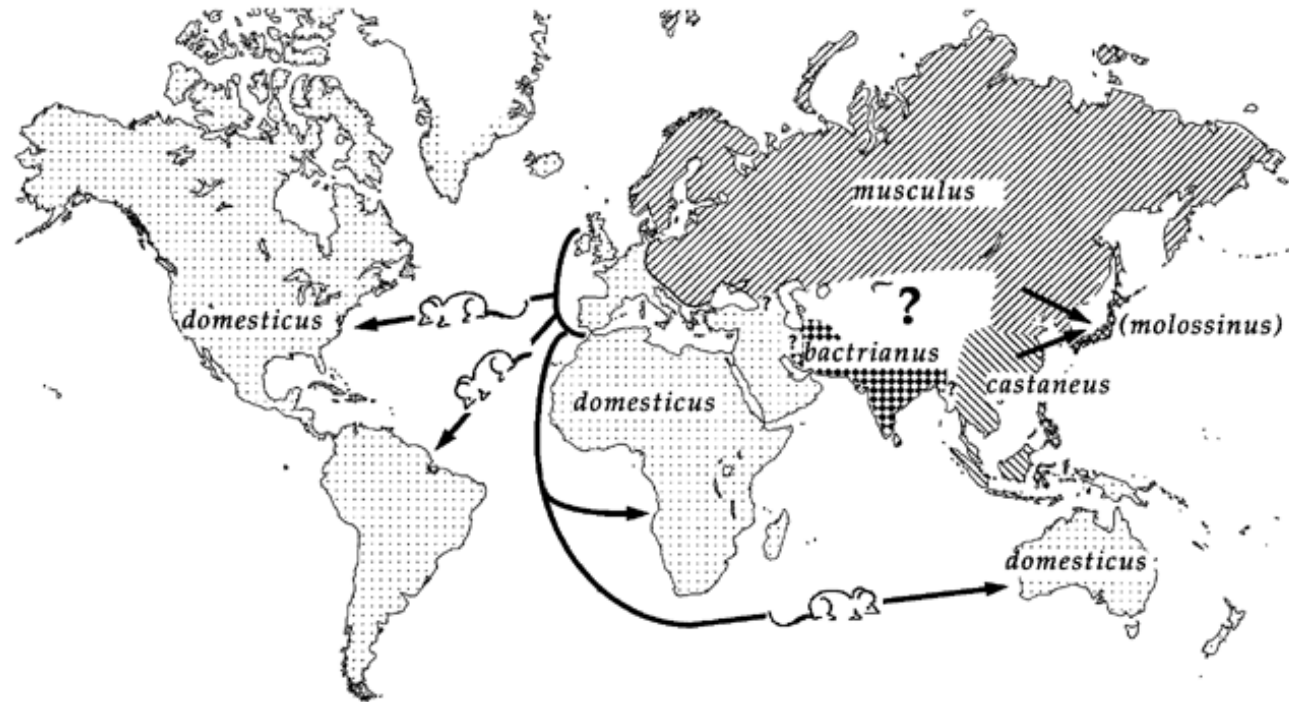# What mice are we using ?
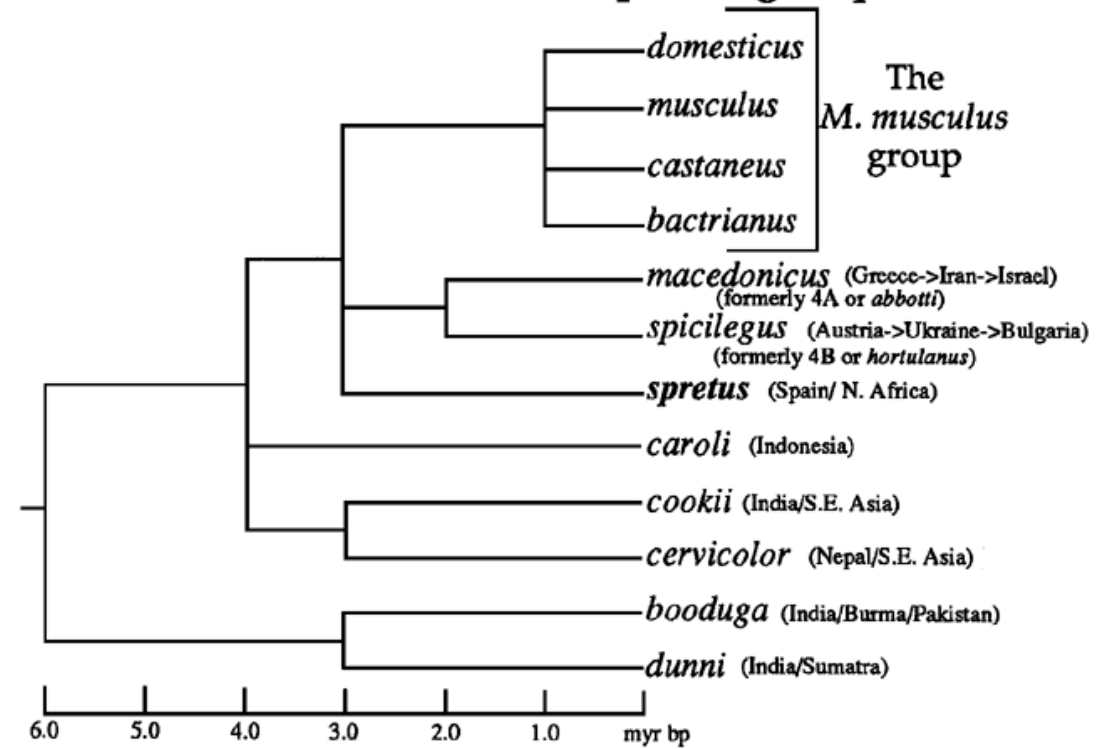# Fancy mice become lab mice

William Ernest Castle (Harvard)

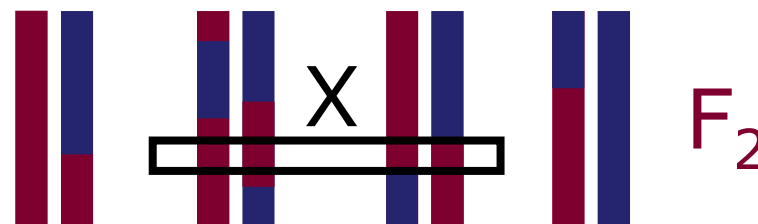Abbie Lathrop, a retired schoolteacher, breeds fancy mice (Granby, MA)

Mus species group

domesticus
musculus — The M. musculus group
castaneus
bactrianus

macedonicus (Greece->Iran->Israel)
(formerly 4A or abbotti)

spicilegus (Austria->Ukraine->Bulgaria)
(formerly 4B or hortulanus)

spretus (Spain/ N. Africa)

caroli (Indonesia)

cookii (India/S.E. Asia)

cervicolor (Nepal/S.E. Asia)

booduga (India/Burma/Pakistan)

dunni (India/Sumatra)

6.0   5.0   4.0   3.0   2.0   1.0   myr bp

Inbred strains: 20 generations of brother x sister matings

X Parental

X F$_1$

X F$_2$
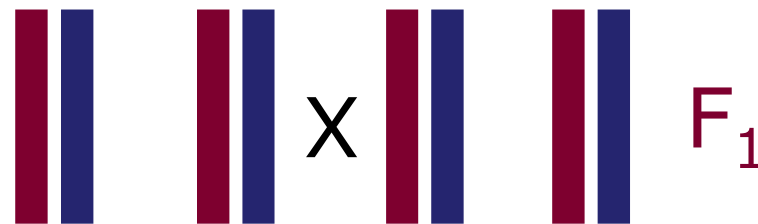
First inbred strain: DBA
bred by Clarence Cook Little (Harvard)
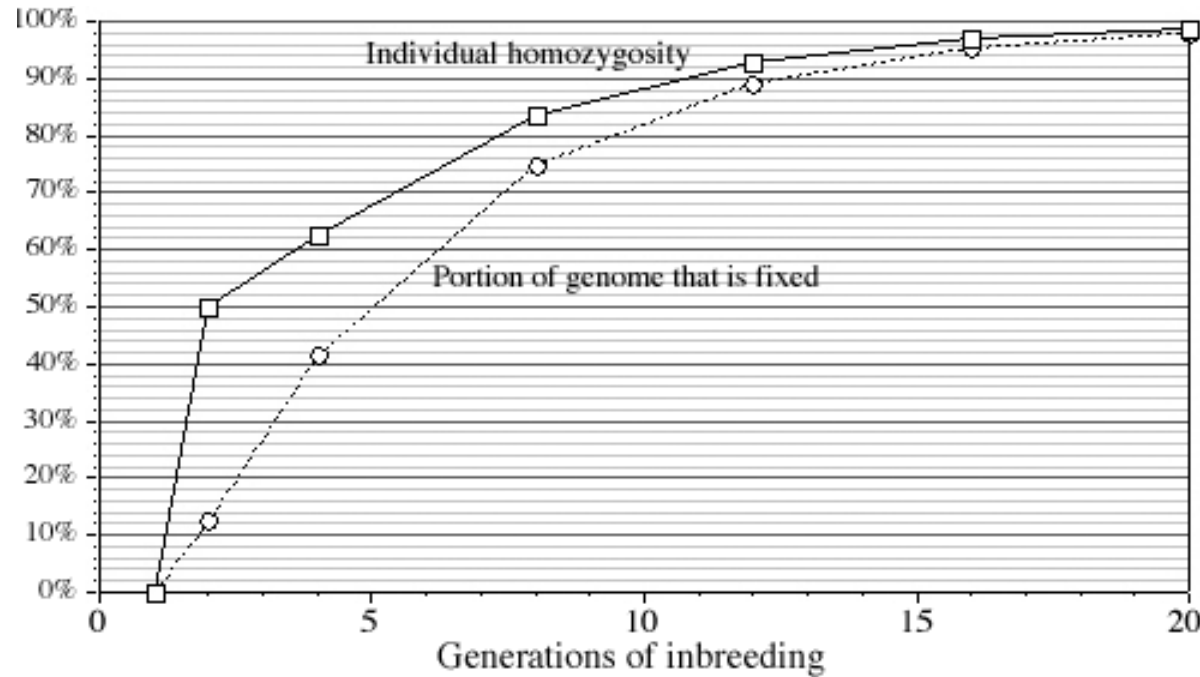
- At F_{20} generations: 98.7% of loci homozygous

Commonly used inbred strains

Strain	Use	Breeding performance	Remarks
129	Construction ES cell lines	Poor	Poor cognition
Balb/c	Hybridoma generation	Poor	Albino
C57BL/6	Reference strain	Good	Old age hearing impaiment
C3H	Common in genetic studies	Poor/Good	Large animals
DBA/2	For F1 and other inbreds	Poor	
FVB	Production of transgenic mice	Excellent	Blind

What's in a mouse name?

- C57BL/6J $\qquad$ C57BL substrain <u>6</u>, <u>J</u>ackson colony

- C57BL/10J $\qquad$ C57BL substrain <u>10,</u> <u>J</u>ackson colony

- C57BL/6NCrl $\qquad$ C57BL/6, <u>N</u>IH colony, distributed by <u>C</u>harles <u>R</u>iver <u>L</u>aboratories

- FVB/NCrl $\qquad$ FVB, <u>N</u>IH colony, distributed by <u>C</u>harles <u>R</u>iver <u>L</u>aboratories

- FVBS/Ant $\qquad$ sighted FVB, <u>Ant</u>werp colony

- F_1: C57BL/6 female x DBA/2 male is named B6D2F1

Lab codes registered with Institute for Laboratory Animal Resources,
USA national acedemy of Sciences, Washington DC

Gene-targeting studies of mammalian behavior: is it the mutation or the background genotype?

Robert Gerlai

Gene targeting to create null mutations in mice is a powerful new tool in biology which will allow the molecular dissection of complex phenotypes such as mammalian brain function, and learning and memory. However, the attempt to interpret the phenotypical changes which arise in null-mutant mice is subject to several caveats. For example, the ability to disrupt a single gene in a targeted manner might lead one to overlook the effects of other genes. Ignoring the genetic background might lead to misinterpretation of results: the present article will demonstrate that the phenotypical abnormalities attributed to the null mutation in several molecular-neurobiological studies could simply result from the effects of background genes.
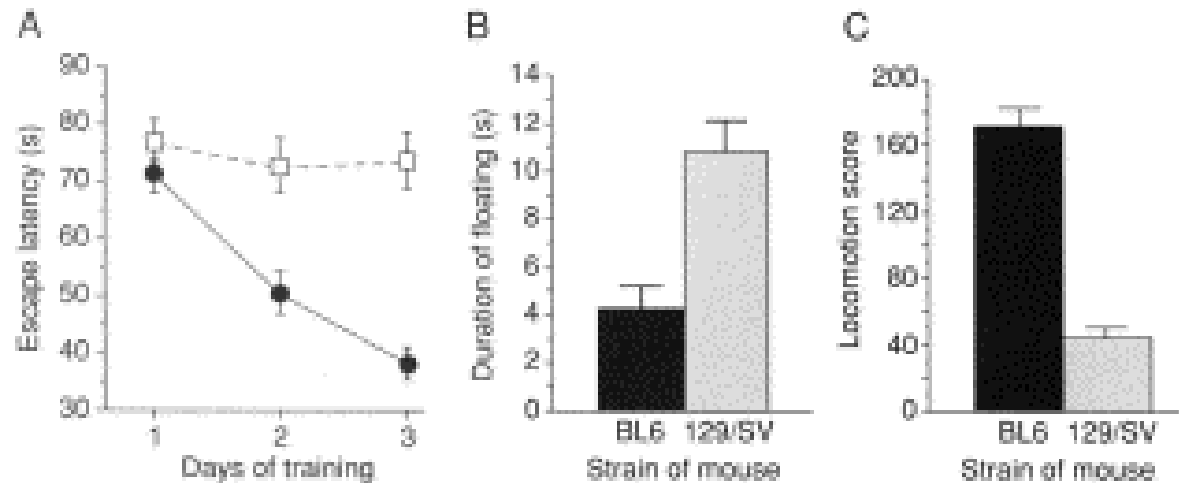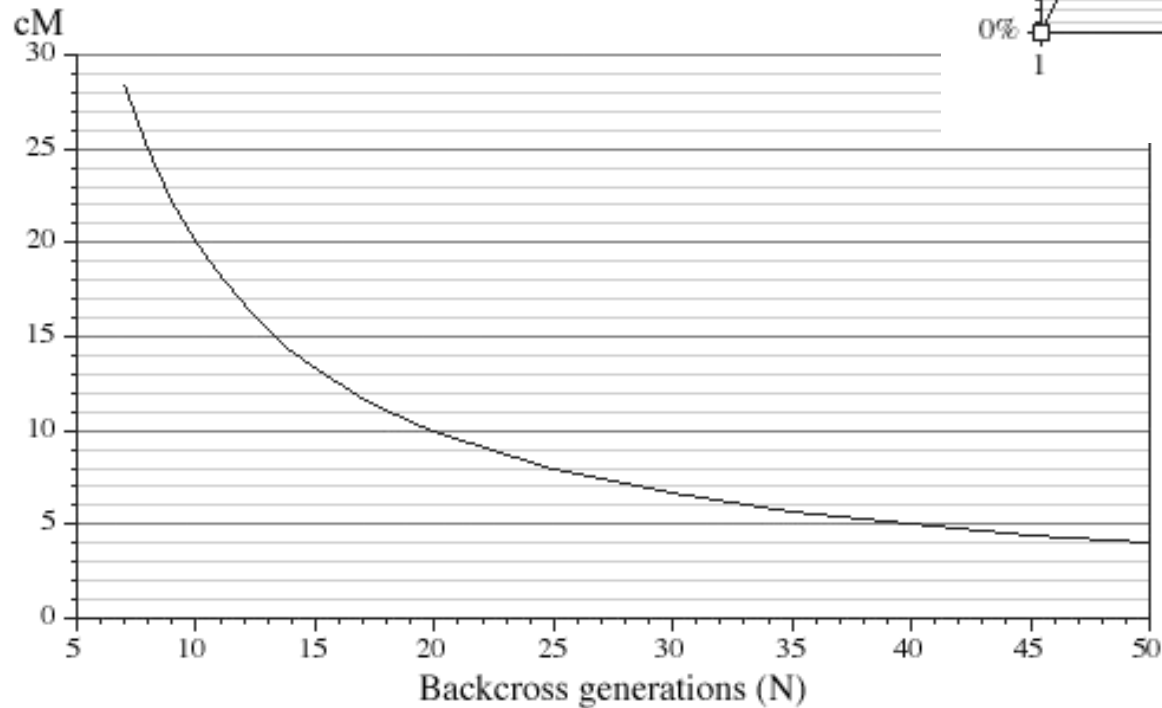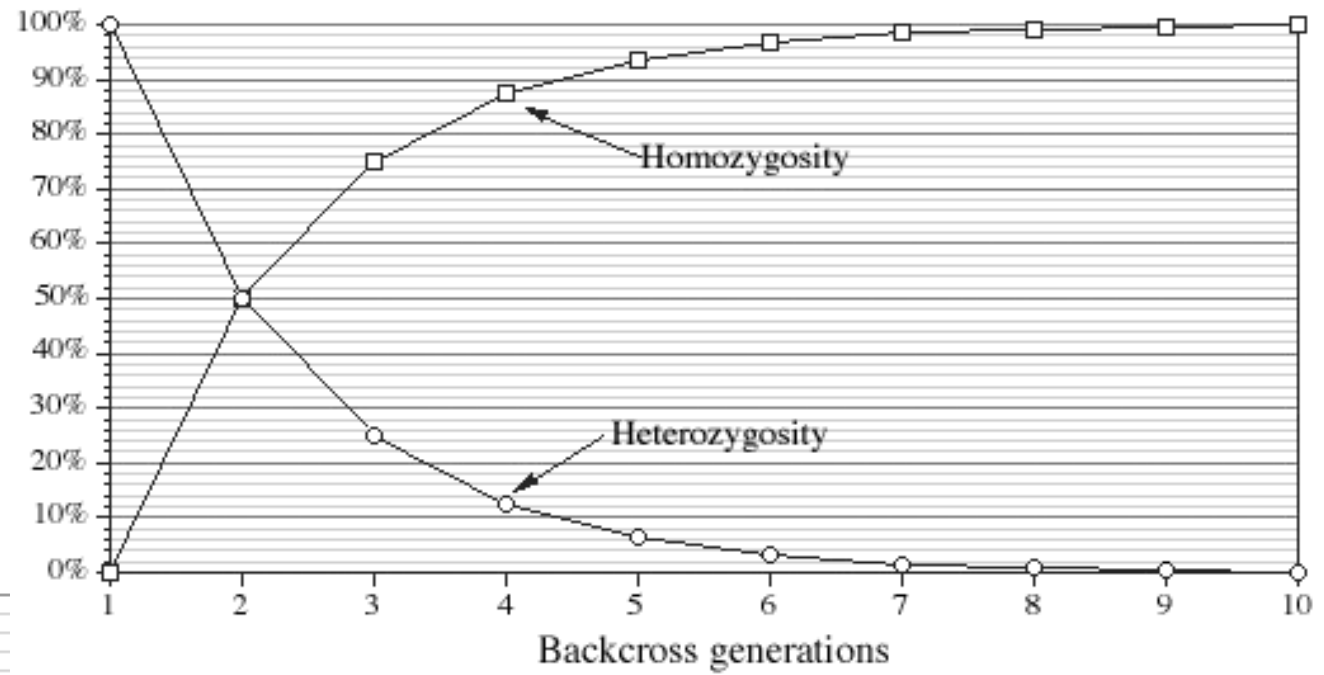
Creation of a congenic strain: 10 repeated backcross matings

Mutation

Parental

F_1
50%

N_2
25%

N_3
75%

N_{4-9}

N_{10}
99.8%
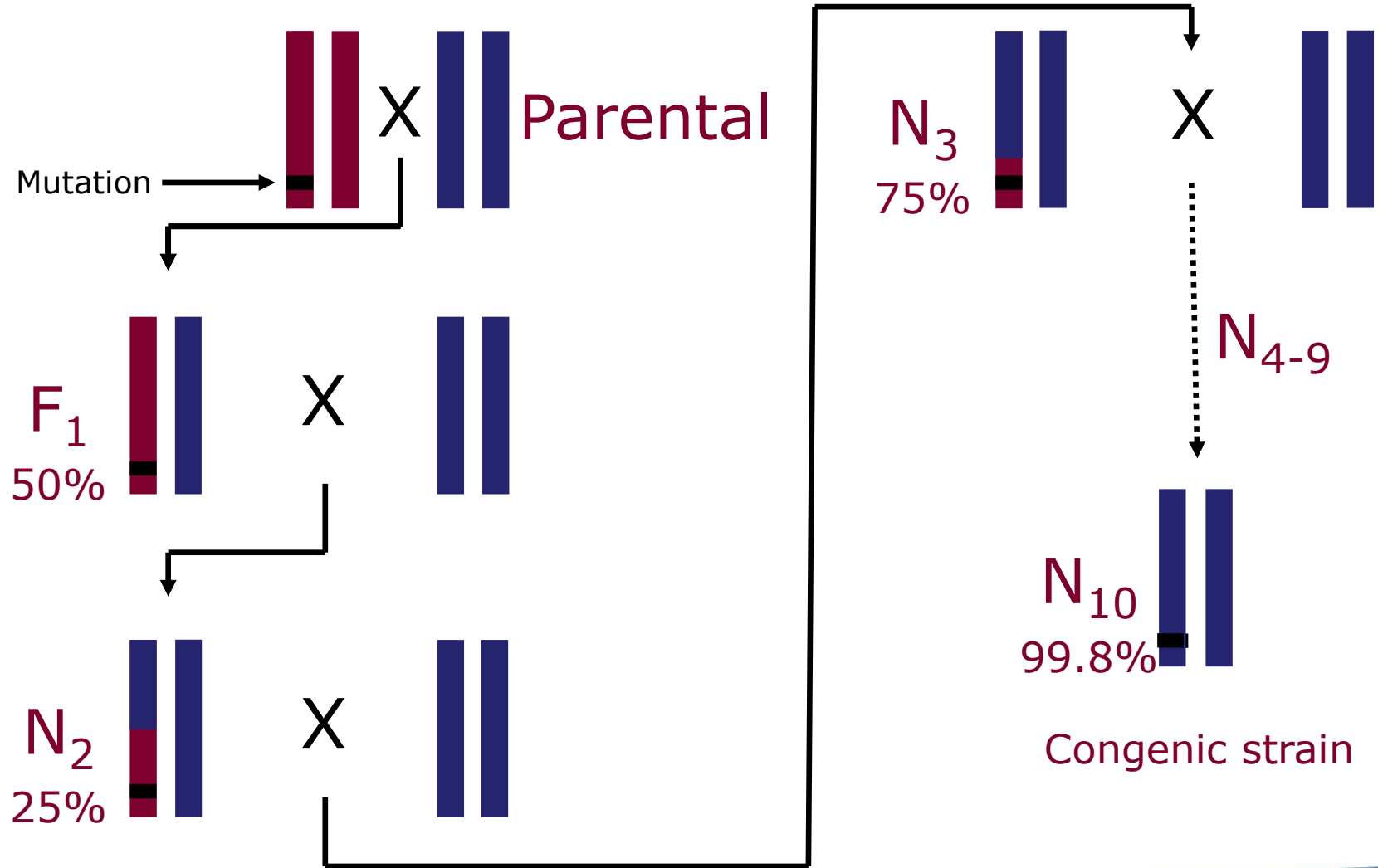
Congenic strain

At generation N_{10}: 98.8% of loci homozygous.......

.... but not at the
the chromosomal segment
flanking the mutation

Genetic nomenclature

- Gene name: *Fmr1 – Italics* and lower case

- Alelles — mutant: *Fmr1*
          — wild type: + or *Fmr1*$^{+}$

- Congenics: abrreviated strain symbols recipient.donor

  B6.129P2 for mutant created in 129P2 backcrossed to C57BL/6 (; if not fully congenic)

- Knockout: congenic name followed by - gene symbol

  B6.129P2-*Fmr1*^{tm1Cgr}/J

1. Tm1 targeted mutation 1
2. Cgr (lab code) created by Ben Oostra, Erasmus University, Rotterdam
3. /J bred at Jackson

2006 The Authors
Journal compilation © 2006 Blackwell Publishing Ltd

Genes, Brain and Behavior (2007)

FVB.129P2-Pde6b$^+$ Tyr$^{c\text{-}ch}$/Ant, a sighted variant of the FVB/N mouse strain suitable for behavioral analysis

V. Errijgers[†], D. Van Dam[‡], I. Gantois[†],
C. J. Van Ginneken[§], A. W. Grossman[¶],
R. D'Hooge[††], P. P. De Deyn[‡,‡‡] and R. F. Kooy[†,*]

[†]Department of Medical Genetics, [‡]Laboratory of Neurochemistry and Behavior, Institute Born-Bunge, Department of Biomedical Sciences and [§]Department of Veterinary Medicine, University of Antwerp, Antwerp, Belgium[¶]Neuroscience Graduate Program, Beckman Institute, University of Illinois at Urbana – Champaign, IL, USA, [††]Department of Biological Psychology, University of Leuven, Leuven, and [‡‡]Memory Clinic, Department of Neurology, Middelheim General Hospital, ZNA, Antwerp, Belgium
*Corresponding author: R. F. Kooy, Department of Medical Genetics, University of Antwerp, Universiteitsplein 1, 2610 Antwerp, Belgium. E-mail: frank.kooy@ua.ac.be

Mice of the FVB/N strain are severely visual impaired as a result of tyrosinase gene defects, leading to a deficiency of the key enzyme for melanin synthesis in skin and eye and of cyclic guanosine monophosphate phosphodiesterase gene defects, which results in albinism (Tyr$^{c/c}$) and retinal degeneration (Pde6b$^{rd1/rd1}$), respectively. Nevertheless, FVB/N mice are commonly used for the generation of transgenic animals because of their large, strong pronuclei and high breeding performance. However, due to visual impairment of the FVB/N animals, the resulting transgenic animals cannot be used in tests that depend on vision, including tests of cognitive behavior. Therefore, we have bred a sighted version of the FVB/N strain by an outcross between FVB/N and 129P2/OlaHsd, followed by repeated backcrosses to FVB/N mice while selecting against albinism and homozygosity of the retinal degeneration mutation. After 11 generations of backcrossing, sighted animals were intercrossed to generate the congenic FVB.129P2-Pde6b$^+$ Tyr$^{c\text{-}ch}$/Ant strain, which is pigmented (Tyr$^{c\text{-}ch/c\text{-}ch}$) and devoid of the genetic predisposition to retinal degeneration. The accurate visual abilities of the FVB.129P2-Pde6b$^+$ Tyr$^{c\text{-}ch}$/Ant mice, for which we propose the name FVBS/Ant, demonstrated a clear visual evoked potential in the presence of normal eye histology and improved performance in the Morris water maze test.
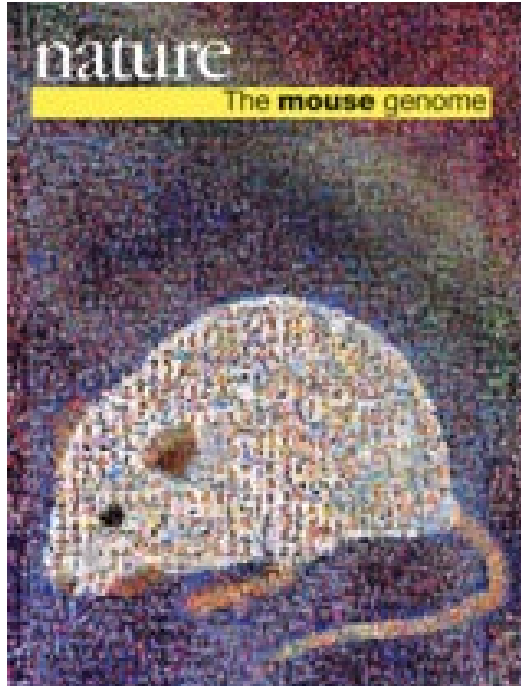

(a) FVB/N   (b) F$_1$ (c) FVBS/Ant

The mouse genome & sequence

2002 Dec
The mouse genome

The Mouse Genome Sequencing Consortium publishes the culmination of international efforts - a high-quality draft sequence and analysis of the genome of the C57BL/6J mouse strain. The estimated size is 2.5 Gb, smaller than the human genome, with less than 30,000 genes. About 40% of the human and mouse genomes can be directly aligned with each other, and about 80% of human genes have one corresponding gene in the mouse genome. Accompanying papers detail various other aspects of the genetic make-up of the mouse, and through the efforts of the RIKEN Genome Science Laboratory in Japan, many essential resources are made freely available.
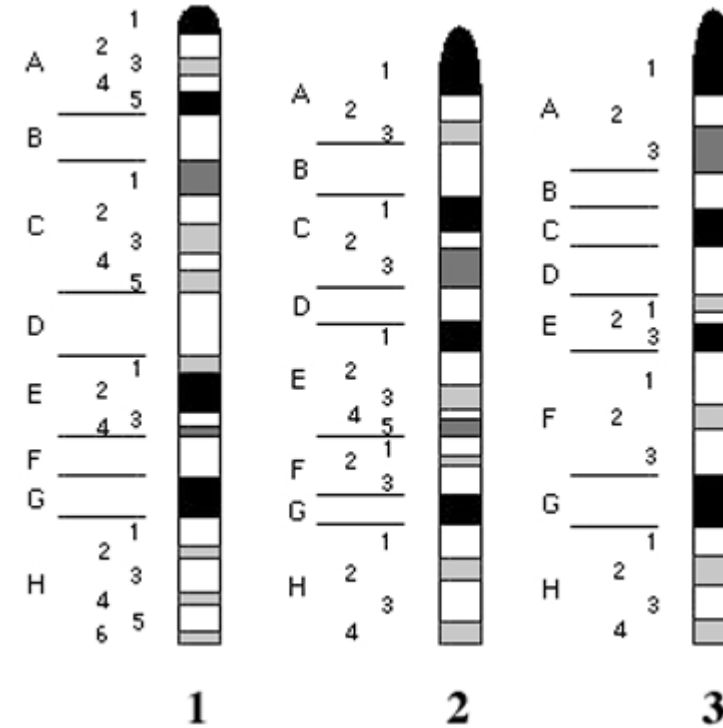
- Genome size: ~2.600 Mb

- Genetic length: ~1450 cM

- Less than 30.000 genes

- Only 1% has no identifiable homologue

- Large set of non-coding RNA
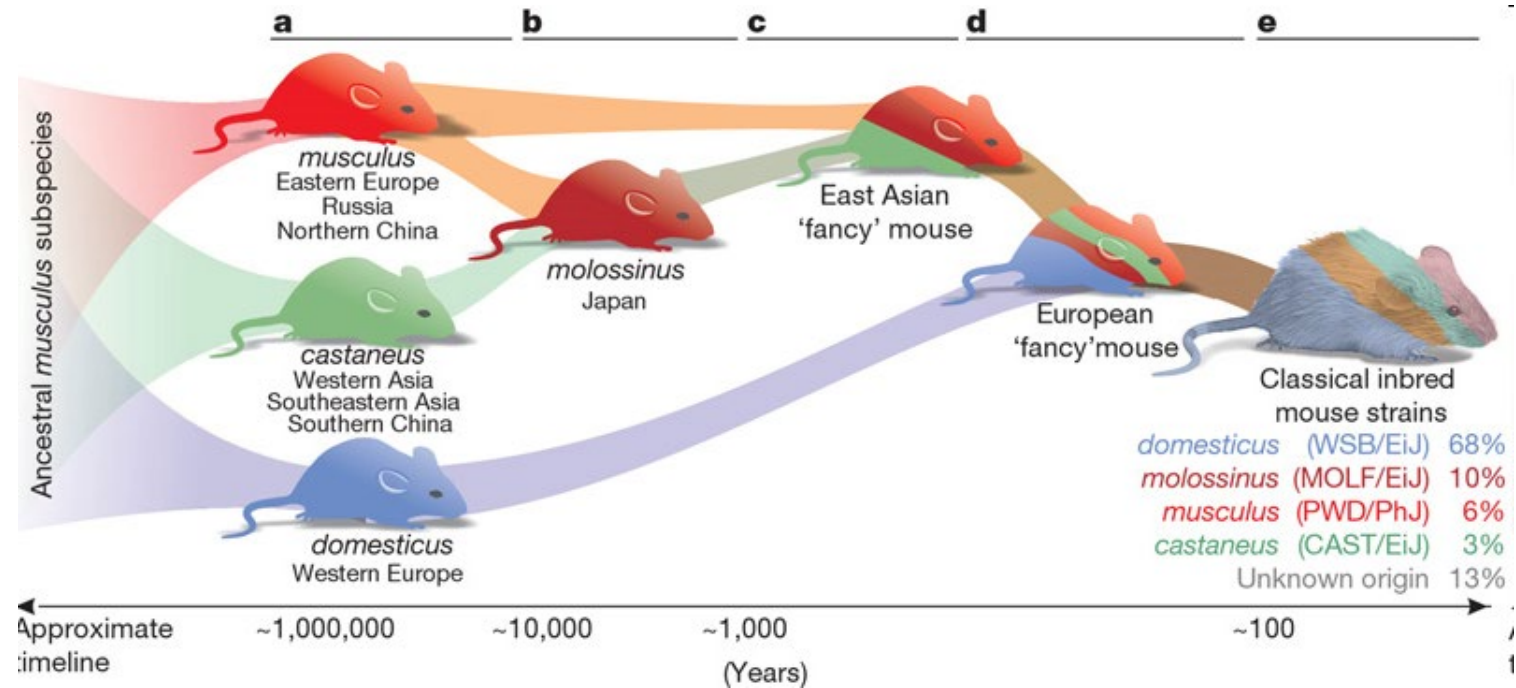
Gap

Mouse karyotype

Mus musculus karyotype

Acrocentric chromosomes,
19 pairs, XY

Laboratory mouse genome

- 1 SNP per 250-300 bp between classical inbred strains
- Mouse Genome consisting of haplotype blocks
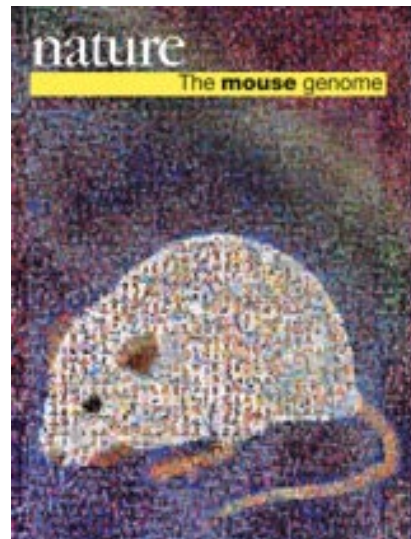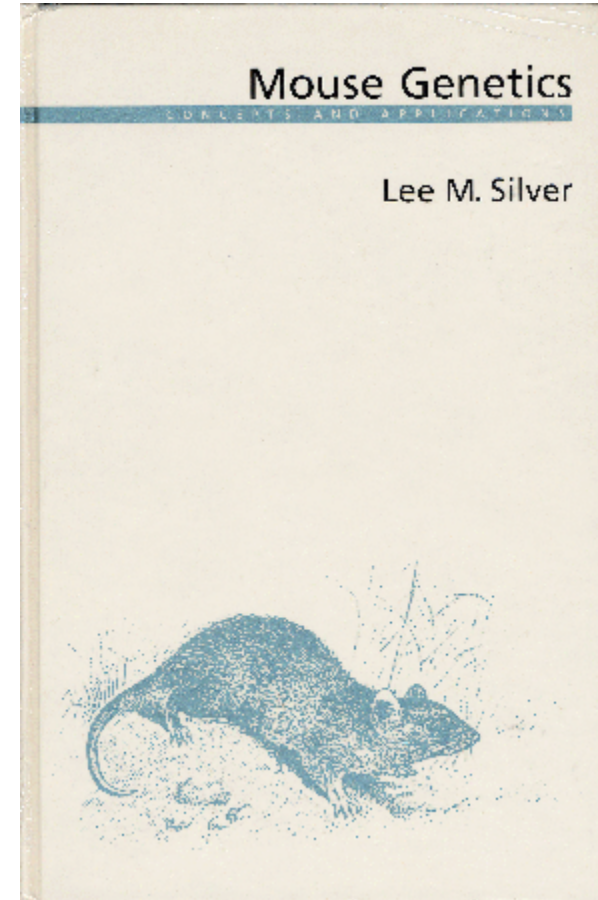- Regions of high and low SNP density exist

Information

http://www.jax.org/index.html

http://www.nature.com/
nature/mousegenome/

Mouse Genetics

Lee M. Silver

http://www.informatics.jax.org/
silverbook/index.shtml