

10 rare tumors that warrant a genetics referral

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Abstract The number of described cancer susceptibility syndromes continues to grow, as does our knowledge on how to manage these syndromes with the aim of early detection and cancer prevention. Oncologists now have greater responsibility to recognize patterns of cancer that warrant referral for a genetics consultation. While some patterns of common cancers are easy to recognize as related to hereditary cancer syndromes, there are a number of rare tumors that are highly associated with cancer syndromes yet are often overlooked given their infrequency. We present a review of ten rare tumors that are strongly associated with hereditary cancer predisposition syndromes: adrenocortical carcinoma, carcinoid tumors, diffuse gastric cancer, fallopian tube/primary peritoneal cancer, leiomyosarcoma, medullary thyroid cancer, paraganglioma/pheochromocytoma, renal cell carcinoma of chromophobe, hybrid oncocytoic, or oncocytoma histol-

ogy, sebaceous carcinoma, and sex cord tumors with annular tubules. This review will serve as a guide for oncologists to assist in the recognition of rare tumors that warrant referral for a genetic consultation.

Keywords Genetic predisposition to disease · Neoplasms · Genetic counseling · Medical genetics · Rare tumor

Introduction

In general, 5–10 % of cancer is due to a hereditary cancer syndrome. Well known examples include hereditary breast and ovarian cancer syndrome and Lynch syndrome. Certain rare tumors are *more* likely to be due to inherited causes. Although rare, these tumors should trigger a cancer genetics work-up in any patient diagnosed with one of these tumors, often regardless of additional personal or family history.

Identification of a hereditary cancer syndrome provides information about future cancer risks for the patient as well as important information for the patient's family members. Knowledge of future cancer risks allows for early detection through targeted high-risk surveillance and in some instances, risk-reducing measures to prevent additional cancers in the patient and family members.

This paper reviews ten rare tumors that have a high likelihood of being due to a hereditary predisposition: adrenocortical carcinoma, carcinoid tumors, diffuse gastric cancer, fallopian tube/primary peritoneal cancer, leiomyosarcoma, medullary thyroid cancer, paraganglioma/pheochromocytoma, renal cell carcinoma of chromophobe, hybrid oncocytoic, or oncocytoma histology, sebaceous carcinoma, and sex cord tumors with annular tubules (Table 1).

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Table 1 Summary of the 10 rare tumors

Associated hereditary cancer syndrome/s	Associated gene/s	Clinical manifestations of hereditary cancer syndrome	% associated with inherited predisposition	Single case indication for referral to genetics (regardless of family history) ^a
<i>Adrenocortical carcinoma</i>				
Li Fraumeni syndrome (LFS)	<i>p53</i>	Adrenocortical carcinoma Sarcomas Breast cancer Leukemia Brain tumors, and others	Up to 80 % of pediatric cases	All individuals with adrenocortical carcinoma
<i>Carcinoid tumors</i>				
Multiple	<i>MEN1</i>	Parathyroid tumors Pituitary tumors Pancreatic tumors	Thyroid gland carcinoid tumors: 25 %	All individuals with thyroid gland carcinoid tumors
Endocrine		Carcinoid tumors of the thymus gland, lung, and stomach	Bronchial and stomach carcinoids: unknown	
Neoplasia type 1 (MEN1)		Other non-endocrine tumors		
<i>Diffuse gastric cancer</i>				
Hereditary diffuse gastric cancer	<i>CDH1</i>	Diffuse gastric cancer Lobular breast cancer	Unknown	All individuals with: Diffuse gastric cancer diagnosed under age 40 Diffuse gastric cancer and lobular breast cancer
<i>Fallopian tube and primary peritoneal cancer</i>				
Hereditary breast and ovarian cancer syndrome	<i>BRCA1</i> <i>BRCA2</i>	Breast cancer (female and male) Ovarian, fallopian tube, and primary peritoneal cancer	Fallopian Tube: 16–43 %	All individuals with fallopian tube or primary peritoneal cancer
<i>Leiomyosarcoma</i>				
HLRCC	<i>FH</i>	Leiomyomas of the skin and uterus Papillary type 2 and collecting duct renal carcinomas Uterine leiomyosarcoma Colorectal cancer Endometrial cancer Gastric cancer Ovarian cancer Hepatobiliary cancer Urinary tract cancer Small bowel cancer Glioblastoma Sebaceous carcinomas	Primary Peritoneal: 28–41 % in Ashkenazi population Unknown	Individuals with: Leiomyosarcoma ND a personal or family history suggestive of HLRCC or Lynch syndrome Multiple cutaneous leiomyomas with at least one histopathologically confirmed leiomyoma
Lynch syndrome (HNPCC)	<i>MSH2</i> <i>MLH1</i> <i>MSH6</i> <i>PMS2</i> <i>EPCAM</i>			

Table 1 continued

Associated hereditary cancer syndrome/s	Associated gene/s	Clinical manifestations of hereditary cancer syndrome	% associated with inherited predisposition	Single case indication for referral to genetics (regardless of family history) ^a
<i>Medullary thyroid carcinoma (MTC)</i>				
MEN2A	<i>RET</i>	MTC Adrenal PC	25 % overall in MTC	All individuals with medullary thyroid cancer
MEN2B	<i>RET</i>	Hyperparathyroidism MTC Adrenal PC		
FMTC	<i>RET</i>	Mucosal neuroma MTC		
<i>Paraganglioma/pheochromocytoma (PGL/PC)</i>				
MEN2	<i>RET</i>	MTC Adrenal PC	>30 % overall in PGL/PC	All individuals with Paraganglioma or Pheochromocytoma
VHL	<i>VHL</i>	Hyperparathyroidism Retinal and CNS hemangioblastoma Renal and pancreatic cysts Renal cell carcinoma Adrenal and extra-adrenal PC Café au lait macules Neurofibromas, Axillary/inguinal freckling, Lisch nodules Adrenal PC		
NFI	<i>NFI</i>			
Hereditary PGL/PC	<i>SDHA</i> <i>SDHB</i> <i>SDHC</i> <i>SDHD</i> <i>SDHAF2</i>	PGL PC Thyroid cancer Kidney cancer GIST		
Familial PC	<i>TMEM127</i> <i>MAX</i>	PC PGL PC PGL		
<i>Renal cell carcinoma—chromophobe, hybrid oncocytotic, oncocytoma histologies</i>				
Birt Hogg Dube	<i>FLCN</i>	Renal carcinoma Follicular hamartomas Lung cysts Spontaneous pneumothorax	Unknown	All individuals with multiple or bilateral chromophobe, oncocytotic, and/or hybrid renal tumors
<i>Sebaceous Carcinomas</i>				
Lynch Syndrome (HNPCC)	See above	See above	50 %	All individuals with a sebaceous carcinoma
<i>Sex cord tumors with annular tubules</i>				

Table 1 continued

Associated hereditary cancer syndrome/s	Associated gene/s	Clinical manifestations of hereditary cancer syndrome	% associated with inherited predisposition	Single case indication for referral to genetics (regardless of family history) ^a
Peutz-Jeghers Syndrome	<i>STK11</i>	Colorectal cancer Gastric cancer Breast cancer Pancreatic cancer Lung cancer Gynecologic cancers	Up to 37 %	All individuals with sex cord tumors with annular tubules

^a The criteria listed in this table are targeted for simplex cases when appropriate. Patients with any of these tumors *and* suggestive family history also meet criteria for referral to genetics

Adrenocortical carcinoma (ACC)

In contrast to benign adrenal cortical tumors (prevalence of 3–5 %), adrenocortical carcinomas (ACCs) are exceedingly rare tumors with an estimated incidence of 0.72 per million in the United States [1]. In the pediatric setting, and to a lesser extent in the adult setting, ACCs are a well known component of at least two genetic conditions: Li-Fraumeni syndrome (LFS) and Beckwith-Wiedemann syndrome (BWS). There have also been reports of an association between ACC and neurofibromatosis type I, multiple endocrine neoplasia type 1, and familial adenomatous polyposis [2–9] but these associations are less established than the two afore-mentioned syndromes.

Li-Fraumeni syndrome

Li-Fraumeni syndrome is a highly penetrant cancer predisposition syndrome associated with pediatric and adult malignancies. It is caused by inherited mutations in the *p53* gene. Individuals with LFS are at risk for a wide range of malignancies, with lifetime cancer risks approaching 100 % for females and 73 % for males [10–12]. Individuals are at risk for multiple primary tumors as well as radiation-induced tumors [13]. The “core” cancers associated with LFS include ACC, sarcomas, breast cancer, leukemia, and brain tumors, however there are many other tumors individuals with LFS are at risk to develop [14, 15]. In families with LFS, the median age at diagnosis of ACC is 3 years, but it can also occur in early adulthood [16].

Diagnostic criteria for classic LFS require a proband with a sarcoma before the age of 45, a first degree relative with cancer before the age of 45, and another first or second degree relative with cancer before the age of 45 or a sarcoma at any age [17]. Studies have found that 50–70 % of families meeting these classic criteria for LFS have an identifiable *p53* mutation [18–22]. In addition to the diagnostic criteria, criteria also exist for consideration of *p53* genetic testing, with lower testing sensitivity [20, 23].

A significant proportion (at least 7 %) of probands with *p53* mutations appear to carry *de novo* mutations [24]. In such cases, family history is often negative. A recent study found that children with ACC may have one of the highest probabilities, up to 80 %, of carrying a *p53* mutation [25]. Per the 2009 Chompret criteria for *p53* testing, all individuals diagnosed with adrenocortical carcinoma at any age, regardless of family history, are candidates for germline *p53* genetic testing [26], preferably in the setting of pre-test genetic counseling.

Management of individuals with LFS must consider the dramatically increased risk for cancer, the early age of onset of these cancers, and the wide spectrum of cancers that can occur. Management is generally impacted by

patient age, personal cancer history and prognosis, and the spectrum of cancers occurring within the family. Historically, there has been limited data to show a particular surveillance protocol can decrease morbidity or mortality in individuals with LFS. However, there is emerging data on a small LFS population that shows frequent biochemical and imaging surveillance may improve survival [27]. In addition, management recommendations for individuals with LFS can also be found in the National Comprehensive Cancer Network (NCCN) Practice Guidelines.

Beckwith-Wiedemann syndrome

Pediatric ACCs are also associated with Beckwith-Wiedemann syndrome (BWS) [28]. A clinical diagnosis of BWS is typically made based on the combination of macroglossia, pre and postnatal overgrowth, and abdominal wall defects (omphalocele, umbilical hernia, or diastasis recti) [29]. Other features of BWS may include hemihypertrophy, kidney anomalies, distinctive ear pits and creases, and an increased risk for a number of malignancies, including ACCs [30]. In contrast to ACC within LFS families, ACC is *not* typically the initial symptom leading to a diagnosis of BWS but rather is a complication to be aware of within patients with BWS.

Carcinoid tumors

Carcinoid tumors represent a heterogeneous group of tumors that arise from diffuse components of the endocrine system. Carcinoids are typically indolent but have malignant potential and comprise less than 1 % of all malignancies. Their overall incidence is estimated at 1–2 per 100,000 individuals. The age distribution of carcinoid tumors ranges from the second to the ninth decade, with the peak incidence occurring between 50 and 70 years of age [31, 32]. More than 60 % of carcinoids originate in the gastrointestinal tract (with half of these in the small intestine) and the remaining in the lungs/bronchi [31].

Carcinoid tumors have been associated several hereditary cancer syndromes, specifically multiple endocrine neoplasia type 1 (MEN1) and neurofibromatosis type 1 (increased risk for periampullary tumors, particularly somatostatin-rich Carcinoids) [33–35]. The most frequent association has been with multiple endocrine neoplasia type 1 (MEN1), specifically carcinoid tumors of the thymus gland, lung and stomach [36–38].

MEN1 is an autosomal dominant syndrome characterized by development of tumors in the parathyroid glands, pituitary and pancreas. It is caused by inactivating mutations of a putative tumor suppressor gene, *MEN1*. MEN1 onset commonly occurs between 15 and 40 years of age

[39]. Hyperparathyroidism is often the first sign of MEN1 and typically occurs between the ages of 20 and 25 years [40]. By age 70, nearly 100 % of MEN1 patients will develop parathyroid tumors, 30–75 % will develop pancreatic endocrine tumors and between 10 and 60 % will develop a pituitary tumor [41]. Non-endocrine tumors are also common in patients with MEN1 and can include lipomas, facial angiofibromas, cutaneous collagenomas, leiomyomas or benign thyroid adenomas and are useful adjunct features in establishing a clinical diagnosis.

Individuals with MEN1 can also develop tumors of the adrenal cortex and carcinoid tumors of the thymus gland, lung or stomach. Thymic, bronchial, and type II gastric enterochromaffin-like (ECL) carcinoids occur in approximately 10 % of individuals with MEN1 syndrome [42, 43]. Typically carcinoid tumors in MEN1 do not cause symptoms until they reach an advanced stage. As these tumors are capable of infiltrating surrounding tissues and metastasizing, early detection is important. Of all the MEN1-related tumors, thymic carcinoids are the most aggressive and management recommendations for patients with MEN1 incorporate surveillance for these tumors.

MEN1 is diagnosed clinically when two of the three major endocrine tumors (parathyroid glands, pituitary and pancreas) are present, or when an individual has one of the major tumors and has a first degree relative with two of the three endocrine tumors, or when an individual harbors an MEN1 disease-causing germline mutation. Germline *MEN1* mutations have been found in 75–90 % of patients with a clinical diagnosis of MEN1, regardless of family history, and approximately 10 % of these are *de novo* mutations. Amongst individuals with thymic carcinoids, approximately 25 % are due to a germline *MEN1* mutation. Given this high prevalence of causative gene mutations, a genetic evaluation for *MEN1* is warranted and recommended for all patients with thymic carcinoid.

Experts have generally suggested initiating MEN1 management in adolescence, but the optimal surveillance initiation age, tests, and frequencies are still undefined. Best clinical judgment guidelines proposed by the International Workshop on MEN1 [41] recommend to start annual biochemical screenings and periodic imaging studies at age 5 years for known or suspected carriers of *MEN1* mutations. Detailed management recommendations for patients with MEN1 can be found in this reference and in the applicable NCCN Practice Guidelines.

Diffuse gastric cancer

Although gastric cancer is one of the most common cancers worldwide, only 1–3 % is due to an inherited syndrome [44, 45]. Hereditary gastric cancers are typically of the

diffuse or linitis plastica histologic type, as opposed to the more common intestinal histologic type [45–47]. Hereditary Diffuse Gastric Cancer (HDGC) is clinically diagnosed in families with either two or more cases of diffuse gastric cancer among first/second degree relatives when one is diagnosed under the age of 50, or families with three cases of diffuse gastric cancer among first/second degree relatives diagnosed at any age [48–50]. In populations with low rates of gastric cancer, 30–50 % of families meeting these criteria have been shown to carry germline *CDH1* mutations [51–53]. *CDH1* (E-cadherin) codes for a highly conserved cell adhesion molecule which is highly expressed in epithelial tissues [54].

Lifetime risk for gastric cancer in patients with HDGC is greater than 80 % and women also have up to a 60 % risk for breast cancer, specifically lobular histology [45, 51, 55]. Emerging data indicates that there may also be an increased risk for colon cancer in families with HDGC [45]. The average age at gastric cancer diagnosis is 38 years but reported ages range from 16 to 68 [53]. Surveillance recommendations for gastric cancer in HDGC have been controversial given limited efficacy of endoscopy for detection of diffuse (intermucosal) gastric cancer [56, 57]. Prophylactic gastrectomy is an option with relatively high uptake per case series of HDGC families; in 2010, the International Gastric Cancer Linkage Consortium recommended that patients with *CDH1* mutations be advised to consider prophylactic gastrectomy [45, 51, 52]. High-risk breast surveillance with inclusion of breast MRI has been recommended for women with HDGC given the elevated risk for lobular breast cancer [45].

While the criteria described above are used to make a clinical diagnosis of HDGC, *CDH1* genetic testing can aid in the evaluation of families suspicious and/or diagnostic of HDGC. A clinical genetic evaluation is available and recommended for any patient meeting the above diagnostic criteria or any one of the following criteria: (1) single case of diffuse gastric cancer diagnosed before the age of 40; (2) both diffuse gastric cancer and lobular breast cancer in the same woman; (3) a family member with diffuse gastric cancer and a family member with lobular breast cancer [45, 58]. Identification of the genetic cause in these families allows for high-risk management in the proband, identifies the elevated risk for breast cancer in women, and allows for identification of family members who are, and are not, at significantly heightened risk for gastric and breast cancer.

Fallopian tube and primary peritoneal cancers

Primary peritoneal carcinomas and fallopian tube carcinomas are rare gynecologic malignancies, often difficult to distinguish pathologically from primary ovarian cancers

[59]. Over the past 10 years, it has become evident that these tumors can be associated with the well-recognized hereditary breast and ovarian cancer (HBOC) syndrome [60–62].

HBOC is caused by inherited mutations in the *BRCA1* and *BRCA2* (*BRCA*) genes. In women, *BRCA* mutations confer a 45–87 % lifetime risk of breast cancer and an 11–54 % risk of ovarian cancer [63–67], compared to general population risks of 12 and 1.4 %, respectively [68]. For women with *BRCA* mutations, the risk ratio to develop a fallopian tube cancer has been calculated to be 11.3 [60] and following prophylactic bilateral salpingo-oophorectomy, the remaining risk of primary peritoneal cancer is estimated to be as high as 4.3 % at 20 years post-BSO [69]. In men, *BRCA2* mutations confer up to a 7 % lifetime risk of breast cancer; the risk is felt to be considerably lower in *BRCA1* mutation carriers [70, 71]. Other cancers in which there has been shown to be an increased risk include prostate cancer, pancreatic cancer, and malignant melanoma [70, 72].

Several small studies, often in isolated ethnic populations, have looked to determine the incidence of *BRCA* mutations in individuals with primary peritoneal carcinomas or fallopian tube carcinomas, unselected for age and family history. In two studies of Ashkenazi Jewish populations in which the carrier frequency of *BRCA* mutations is increased, 28–41 % of women with primary peritoneal carcinoma were found to harbor a founder *BRCA* mutation [60, 73]. In a Canadian sample of 51 women with fallopian tube cancers, 16 % were found to carry *BRCA* mutations [74]. In an American sample of 28 patients with primary fallopian tube cancers, 43 % were found to carry *BRCA* mutations [75]. In a study of 29 Ashkenazi Jewish women with primary fallopian tube cancers, 17 % were found to carry a *BRCA* founder mutation [60]. For both types of cancer, the diagnosis has been shown to occur on average 8–10 years younger in *BRCA* mutation carriers in comparison to those with sporadic cancers [60, 75].

A variety of risk management options exist for mutation carriers, including increased surveillance, chemoprevention, and prophylactic surgical options. Detailed management recommendations for individuals with HBOC can be found in the NCCN Practice Guidelines. Given the risk for fallopian tube cancer, it has been recommended that female *BRCA* mutation carriers who undergo risk-reducing oophorectomy have their fallopian tubes removed as well [61, 62, 76]. Given the risk of an occult cancer at the time of prophylactic bilateral salpingo-oophorectomy (PBSO), serial-sectioning of the ovaries and fallopian tubes is recommended following a high-risk protocol [76–78]. Studies have shown that 2–17 % of *BRCA* mutation carriers who undergoing PBSO are found to have an occult ovarian or fallopian tube carcinoma at the time of surgery, many of

which are shown to arise in the fimbriae of the fallopian tube [69, 77, 78].

Primary peritoneal carcinoma and fallopian tube carcinomas are recognized as part of the HBOC tumor spectrum and genetic risk assessment is recommended for any woman with a diagnosis of one of these tumors, regardless of age at diagnosis or family history [79].

Leiomyosarcoma

Leiomyosarcomas (LMS) are malignant mesenchymal tumors with features of smooth muscle differentiation. They account for 23.7 % of all soft tissue sarcomas (STS) and are the most common STS subtype [80]. LMS develop principally in adults between 50 and 60 years of age, are more common in women than men, and can occur in a variety of anatomical sites including the uterus, retroperitoneum, skin, superficial soft tissues, and deep compartments of the extremities [81].

While most LMS are sporadic in origin, LMS have been reported in families with Hereditary Leiomyomatosis and Renal Cell Cancer (HLRCC) Syndrome, Lynch Syndrome, and Hereditary Retinoblastoma. There have also been a few case reports in patients with *p53* and *BRCA* gene mutations [82–84].

Hereditary leiomyomatosis and renal cell cancer (HLRCC) syndrome

This rare syndrome is characterized by leiomyomas of the skin and uterus and renal cell carcinomas (papillary type II and collecting duct) [85]. HLRCC is caused by germline mutations in fumarate hydratase (*FH*), a tumor suppressor gene that encodes for a Krebs cycle enzyme. Mutations are found in approximately 90 % of individuals with cutaneous leiomyomas [86, 87].

In females, the risk for uterine leiomyomas by age 45 years is greater than 75 %, and the risk for cutaneous leiomyomas is greater than 70 % [88]. Nearly all males with HLRCC have cutaneous leiomyomas by age 35 years [85]. The estimated frequency of renal cell carcinoma (papillary type 2 and collecting duct) is approximately 15–30 % [86]. Individuals with HLRCC are also at risk for developing leiomyosarcomas at various sites, most often the uterus [89]. To date, six cases of “uterine leiomyosarcoma” have been reported in HLRCC families; however the exact risk in HLRCC remains unclear [90].

There are currently no consensus criteria for a clinical diagnosis of HLRCC. The diagnosis is generally based on testing the enzymatic levels of fumarate hydratase and/or germline genetic testing. The following features warrant referral for genetic risk assessment: multiple cutaneous

leiomyomas of which at least one has been histopathologically confirmed; an individual who has a single leiomyoma in the presence of a family history of HLRCC; or, an individual who has one or more tubulopapillary, collecting duct, or papillary type II renal tumors, with or without a family history of HLRCC [91].

Identification of HLRCC identifies the increased risk for renal cancer and allows for high-risk management in the proband and family members. At present there is no consensus on clinical surveillance of individuals with HLRCC. Published management recommendations [92, 93] include: baseline renal ultrasound examination and abdominal CT scan with contrast or MRI at age 20 to screen for renal tumors followed by annual MRI and semi-annual ultrasound examinations, annual gynecological ultrasound examination of females starting at age 20, in addition to detailed dermatologic examination for evaluation of lesions suspicious for cutaneous leiomyosarcoma.

Lynch syndrome (also known as hereditary non-polyposis colorectal cancer syndrome, HNPCC)

Lynch syndrome is an autosomal dominant syndrome characterized by an increased risk of colorectal cancer as well as cancers of the endometrium, biliary tract, ovary, ureter and renal pelvis, stomach, pancreas, brain, and sebaceous neoplasias [94]. The lifetime risk of cancer is as high as 80 % by age 70 with colon and uterine being the most common tumors [95, 96]. Lynch Syndrome is caused by an inherited mutation in the DNA mismatch repair genes *MLH1* and *MSH2* and, to a lesser extent, *MSH6*, *PMS2*, and *EPCAM (TACSTD1)* [97, 98].

In 2002, Medina et al. [99] reported the occurrence of a pleomorphic paravertebral leiomyosarcoma in a young female whose family history was diagnostic of Lynch syndrome. Tumor analysis revealed microsatellite instability, reinforcing the association of the leiomyosarcoma as a component Lynch tumor in this family. Additional cases of leiomyosarcomas in individuals with Lynch Syndrome have been reported [100, 101]. While not formally part of Lynch Syndrome, individuals with LMS should be worked up for Lynch Syndrome in the context of additional suggestive personal and/or family history. Identification of Lynch Syndrome allows for implementation of high risk surveillance and prevention options to reduce morbidity and mortality. Management recommendations for patients with Lynch Syndrome can be found in the NCCN Practice Guidelines.

Hereditary retinoblastoma

Long-term follow-up of a cohort of survivors of hereditary retinoblastoma revealed a statistically significant excess of

leiomyosarcoma and other soft tissue sarcomas that persists decades after the retinoblastoma (RB) diagnosis [102]. Osteosarcoma is the most common soft tissue secondary malignancy in individuals with hereditary retinoblastoma, although there have been at least 18 cases of leiomyosarcoma reported as a secondary neoplasm [103]. These often originate within the field of prior radiation with 78 % of leiomyosarcomas diagnosed 30 or more years after the retinoblastoma diagnosis. Bladder leiomyosarcoma has also been associated with hereditary RB with the longest reported interval between diagnosis and follow-up evaluation being 5 years [102, 104].

At this point in time, a single case of leiomyosarcoma does *not* warrant genetic testing for a particular gene/s but if a detailed personal and family history identified features suggestive of one of the aforementioned hereditary cancer syndromes, genetic testing would be indicated [92]. Genetic counseling for patients with leiomyosarcoma can help determine which cases warrant genetic testing and which do not.

Medullary thyroid carcinoma

Medullary thyroid carcinoma (MTC) accounts for 5–10 % of all thyroid cancers diagnosed in the U.S. [105]. Twenty-five percent of these cases are hereditary (due to germline *RET* mutations) [106]. Germline mutations in the *RET* proto-oncogene cause Multiple Endocrine Neoplasia Type 2 (MEN2). MEN2 can be further categorized into MEN2A, MEN2B, and Familial Medullary Thyroid Carcinoma (FMTC). All MEN2 subtypes confer a nearly 100 % lifetime risk of MTC. Individuals with MEN2A also have up to a 50 % risk of pheochromocytoma and up to a 20–30 % risk of hyperparathyroidism. Those with MEN2B have up to a 50 % risk of pheochromocytoma and exhibit characteristic physical features, including a Marfanoid habitus and mucosal neuromas [107]. FMTC has historically been used to describe families in which four or more individuals have MTC in the absence of pheochromocytoma or hyperparathyroidism [108]. Some individuals with MEN2A or FMTC may also have Hirschsprung's disease due to a specific mutation in exon 10 of the *RET* gene [108].

Genetic testing for the *RET* proto-oncogene is available through several clinical laboratories. Testing typically begins with select exons (10, 11, 13–16), and if no mutation is identified, more extensive testing may be considered. In patients with a clinical diagnosis of MEN2, over 98 % will have an identifiable *RET* mutation [108]. For those presenting with “sporadic MTC,” that is MTC in the absence of family history or other signs of MEN2, approximately 7 % will be found to have a germline *RET* mutation [109].

Genetic testing is not only a highly sensitive method used to diagnose MEN2, it also helps to direct medical management as significant genotype-phenotype correlations exist [106]. Proband presenting with MTC should undergo standard surgical treatment and surveillance for residual or recurrent disease [106, 110]. Prophylactic thyroidectomy is recommended for at-risk family members who test positive for a familial mutation. The age at which to perform prophylactic thyroidectomy or biochemical screening for MTC, as well as the age to begin surveillance for pheochromocytoma and/or hyperparathyroidism is dependent on the specific *RET* mutation identified [106]. In contrast to many hereditary cancer syndromes, genetic testing for children *is* recommended because of the obvious benefits of prophylactic thyroidectomy and early surveillance [106].

Given that 25 % of MTC cases are hereditary, all individuals with a history of MTC should be offered genetic testing in the setting of genetic counseling [106].

Paranglioma and pheochromocytoma

Parangliomas and pheochromocytomas are rare tumors derived from the parasympathetic and sympathetic nervous systems [111]. Parasympathetic head and neck parangliomas are typically non-functioning. The sympathetic parangliomas arise from the adrenal medulla or extra-adrenal ganglia and are typically functionally active as indicated by excess catecholamines [112]. The term pheochromocytoma is used to describe adrenal, intraabdominal, and thoracic catecholamine-producing parangliomas.

Parangliomas/pheochromocytomas are a key component of the hereditary cancer syndromes described below. Approximately 30 % of all individuals with a paranglioma or pheochromocytoma harbor a germline mutation in *RET*, *VHL*, *SDHB*, *SDHC*, *SDHD*, or *NF1* [113]. Four additional susceptibility genes, *SDHAF2*, *TMEM127*, *SDHA*, and *MAX* have recently been identified which indicates an even higher prevalence of hereditary predisposition to these tumors [114, 115] [116, 117].

Multiple endocrine neoplasia type 2 (MEN 2)

As described in the previous section, MEN2, caused by mutations in the *RET* proto-oncogene, is a hereditary condition which increases the risk for MTC and pheochromocytoma, as well as primary hyperparathyroidism in individuals with MEN2A. While most patients with MEN2 present with MTC, pheochromocytomas are the first sign in 13–27 % of individuals with MEN2A [118, 119]. Pheochromocytomas in patients with MEN2A are diagnosed at an earlier age, have subtler symptoms, and are more likely

to be bilateral than sporadic tumors [120, 121]. Identifying MEN2 in a patient with pheochromocytoma who has not yet been diagnosed with MTC allows for prophylactic, or in many cases, therapeutic, thyroidectomy.

Von Hippel-Lindau syndrome (VHL)

Germline mutations in the *VHL* tumor suppressor gene cause von Hippel-Lindau Syndrome (VHL). This hereditary cancer syndrome is characterized by hemangioblastomas of the retina, spine, and brain stem; renal cysts and clear cell renal cell cancer; pancreatic cysts and pancreatic islet cell tumors; endolymphatic sac tumors; epididymal cysts; and adrenal or extra-adrenal pheochromocytomas. Approximately 20–30 % of individuals with VHL are diagnosed with pheochromocytoma, typically at an early age (average 22 years) [122]. Genotype/phenotype correlations have been described, with missense mutations leading to higher risks of pheochromocytomas, and a lower incidence seen in families with large deletions or truncating mutations [123]. Identification of VHL in patients presenting with pheochromocytoma allows for appropriate medical management and surveillance including ophthalmologic screening and imaging of the abdomen, spine, and brain to identify benign and malignant lesions.

Hereditary paraganglioma-pheochromocytoma syndromes (PGL/PC)

Mutations of the *SDHB*, *SDHC*, and *SDHD* genes are associated with the three hereditary PGL/PC syndromes known as PGL 4, PGL 3, and PGL 1. Recently, two additional genes have been added to this syndrome. The gene associated with PGL 2 was identified as *SDHAF2* and mutations in the *SDHA* gene were proven to be associated with pheochromocytoma [116, 124]. Variation in the common locations and nature of tumors exists amongst the PGL/PC syndromes; *SDHB* mutation carriers are more likely to be diagnosed with extra-adrenal pheochromocytomas and have a higher risk of malignancy, while benign head and neck paragangliomas predominate in *SDHD*, *SDHC*, and *SDHAF2* mutation carriers [114, 125, 126]. There have been reports of gastrointestinal stromal tumors in individuals with hereditary PGL/PC, as well as renal cell carcinoma and papillary thyroid carcinoma in *SDHB* mutation carriers [127–129]. While no consensus for management of individuals with hereditary PGL/PC exists, yearly biochemical screening and imaging of the head, neck, thorax, abdomen, and pelvis to detect new tumors may be considered [130].

Neurofibromatosis type 1 (NF1)

Adrenal pheochromocytomas arise in less than 6 % of individuals with a diagnosis of NF1 [131]. Clinical

diagnosis of NF1 in adults is based on meeting two of the following criteria: ≥ 6 café-au-lait macules, ≥ 2 neurofibromas or ≥ 1 plexiform neurofibroma, axillary or inguinal freckling, optic glioma, ≥ 2 Lisch nodules, osseous lesions, and positive family history [132]. Genetic testing for *NF1* is not indicated in simplex cases of pheochromocytoma as these individuals may be diagnosed based on clinical findings [133].

Familial pheochromocytoma (FP)

The first report of mutations in *TMEM127* found that one-third of patients with familial pheochromocytoma and 3 % of patients with sporadic pheochromocytoma without a mutation in the previously described pheochromocytoma genes carried a mutation in *TMEM127* [115]. Additional studies have confirmed the finding of *TMEM127* mutations in patients with adrenal pheochromocytoma as well as extra-adrenal and head/neck paraganglioma [134, 135]. Although knowledge of the penetrance of paraganglioma and pheochromocytoma in individuals with *TMEM127* mutations is in its infancy, studies suggest a surveillance approach similar to that in Hereditary Paraganglioma syndrome [134, 136].

Max

MAX is the most recently described hereditary pheochromocytoma and paraganglioma gene. In preliminary studies, it appears as though individuals with mutations in *MAX* have an increased likelihood of developing malignant pheochromocytomas and paragangliomas and tend to have a positive family history and/or multifocal tumors [117, 137]. As with *TMEM127*, more research is necessary to determine the best surveillance approach for individuals with *MAX* mutations.

All of the above syndromes exhibit an autosomal dominant pattern of inheritance. *SDHD*, *SDHAF2*, and *MAX* are maternally imprinted; therefore, children of females with a mutation will not develop paragangliomas despite inheriting the mutation.

Given that at least 30 % of all individuals with a paraganglioma or pheochromocytoma harbor a germline mutation in one of these genes, all patients with this diagnosis should have a thorough genetics evaluation [113]. Factors that increase the likelihood of detecting a germline mutation include multifocal and bilateral tumors, early age of onset (<45 years), positive family history, and extra-adrenal location [133]. A genetics professional can help to prioritize the gene(s) to be tested by evaluating the patient for features of the associated hereditary conditions.

Renal cell carcinoma—chromophobe, hybrid oncocytotic, oncocytoma histologies

Chromophobe, hybrid oncocytotic, and oncocytoma renal cell carcinomas account for less than 10 % of all renal cell carcinomas [46]. These rare tumors are a key component of Birt-Hogg-Dube (BHD), a hereditary genodermatosis syndrome. Diagnosis of this syndrome amongst renal cell carcinoma patients with these histologies influences surgical management, identifies other symptoms to be screened for, and allows for targeted surveillance and intervention in family members.

BHD is an autosomal dominant syndrome caused by mutations in the tumor suppressor gene *BHD* (also known as *FLCN*) [138, 139] and is characterized by follicular hamartomas (fibrofolliculomas), lung cysts, spontaneous pneumothorax, and renal carcinoma [140, 141]. Germline *BHD* mutations have been identified in approximately 85–90 % of patients with histories highly suggestive of BHD [140, 141].

Confirmed fibrofolliculomas have been reported in 70–85 % of individuals with BHD [140–142]. Angiofibromas, trichodiscomas, and perifollicular fibromas have also been reported in BHD families [140]. Cutaneous manifestations are typically distributed over the face, neck, and/or upper trunk [140, 142, 143]. In regards to lung manifestations, approximately 75–90 % of patients develop lung cysts and 33–38 % have a history of spontaneous pneumothorax [140].

Kidney tumors have been reported in 7–34 % of patients with BHD [140]. The prevalence variability likely reflects ascertainment bias as some reports are based on families ascertained by dermatologic findings while others are based on families recruited because of renal tumor histories. Reported age at diagnosis ranges from age 20 to 74 [143]. Renal tumors in BHD patients are typically multifocal and bilateral and are often slow-growing and with mixed inter- and intra-tumor histology [142, 143]. Typical histologies include hybrid oncocytotic (67 %), chromophobe (23 %), and oncocytotic (3 %) although other renal cell carcinoma histologies, including clear cell, have been reported [142].

There are no consensus-based recommendations for management of patients with BHD however expert opinion recommendations have been put forth [142, 143]. Proposed recommendations include routine surveillance for renal tumors beginning between age 20 and 25 [143]. Given the typical nature of renal tumors in patients with BHD, observation of small tumors is recommended, reserving surgery for tumors over 3 cm or tumors demonstrating accelerated growth [142]. Additionally, nephron-sparing surgery is recommended as opposed to radical nephrectomy whenever possible for BHD patients to reduce

morbidity [142, 143]. Identification of BHD in the patient with these rare tumor histologies influences surgical management and also alerts the medical team to the significantly elevated risk of spontaneous pneumothorax.

A genetics evaluation for BHD is recommended for patients with multiple and bilateral chromophobe, oncocytic, and/or hybrid renal tumors and for patients with a single oncocytic, chromophobe, or oncocytic hybrid renal tumor and a family history of renal cancer of these histologies [142–144].

Sebaceous carcinomas

Sebaceous carcinomas are rare adnexal tumors characterized by sebocytic differentiation [145]. These tumors may arise sporadically or in the setting of Lynch syndrome. As described above, Lynch syndrome is inherited in an autosomal dominant pattern due to germline mutations in any one of at least five genes involved in the mismatch repair pathway, *MLH1*, *MSH2*, *MSH6*, *PMS2*, and *EPCAM*. Cancers arising due to defective mismatch repair, including sebaceous carcinomas, exhibit a “molecular fingerprint” known as microsatellite instability [101, 146–151].

Lynch syndrome is characterized by an increased risk for early onset colorectal and endometrial cancers, as well as ovarian, small intestine, stomach, upper urinary tract, and biliary tract cancers and sebaceous neoplasias [152]. When looked at collectively, sebaceous adenomas, epitheliomas, and carcinomas occur in approximately 1–9 % of individuals with Lynch syndrome [153, 154].

The subset of Lynch syndrome characterized by visceral malignancies and sebaceous gland neoplasms and/or keratoacanthomas is also referred to as Muir-Torre syndrome (MTS). Historically, a clinical diagnosis of MTS has been based on the presence of at least one sebaceous neoplasm (sebaceous adenoma or carcinoma) or keratoacanthoma and a visceral malignancy [155]. Now, MTS can be diagnosed in patients with a sebaceous carcinoma *prior* to development of a visceral malignancy via germline genetic testing of the Lynch syndrome-associated genes.

The diagnosis of a sebaceous carcinoma provides an important diagnostic clue to a diagnosis of Lynch syndrome. The percentage of unselected sebaceous carcinomas that arise in the setting of Lynch syndrome is estimated to be at least 50 % [156, 157]. The location of sebaceous carcinomas may also be important when determining a risk of Lynch syndrome, with the highest association found in those occurring in non-head and neck locations [158, 159].

Recognition of Lynch syndrome has important clinical implications for the management of both the index patient and relatives. The benefit of identifying individuals with Lynch syndrome is realized through the initiation of a

heightened cancer surveillance for the index patient and at risk relatives. By enabling early detection, regular colonoscopies reduce the colorectal cancer risk by greater than 50 % in at-risk members of Lynch syndrome families [160]. Detailed management recommendations for individuals with Lynch syndrome can be found in the NCCN Practice Guidelines.

Due to the strong association between sebaceous carcinomas and Lynch syndrome, the uncommon occurrence of sebaceous carcinomas in the general population, and the proven clinical utility of the identification of families with Lynch syndrome, several experts recommend that the diagnosis of a sebaceous carcinoma alone should prompt an evaluation for Lynch syndrome regardless of family history, age of onset, or the presence of other malignancies [153, 161, 162].

Sex cord tumors with annular tubules

Sex cord tumors with annular tubules (SCTAT) are rare tumors that fall within the designation of ovarian sex cord-stromal tumors (OSCST), a heterogeneous group of tumors that develop from the gonadal non-germ cell component of the ovary [163]. SCTATs were first described in 1970 by pathologist, Dr. Robert Scully [164]. In the initial report, he described 10 cases of distinctive-appearing ovarian tumors characterized by simple and complex ring-like tubules and a tendency for calcification. He also noted that three patients had clinical manifestations of Peutz-Jeghers syndrome (PJS), a hereditary polyposis and cancer predisposition syndrome described below [164].

PJS is an autosomal dominant syndrome characterized by hamartomatous polyps of the gastrointestinal tract and by distinctive mucocutaneous hyperpigmentation [165]. A clinical diagnosis of PJS is made when an individual has either: (1) two or more Peutz-Jeghers polyps in the gastrointestinal tract; or (2) one Peutz-Jeghers polyp in the gastrointestinal tract, together with either classic PJS hyperpigmentation or a family history of PJS [166].

In addition to the findings of GI polyposis and mucocutaneous hyperpigmentation, individuals with PJS are predisposed to many cancers, including GI malignancies, breast cancer, pancreatic cancer, lung cancer, testicular cancer, and gynecologic malignancies [167–172]. The lifetime risk of cancer is as high as 67–93 % by ages 65–70 [168, 170–172]. Identification of PJS is imperative given the significant increases in cancer risk and the availability of targeted surveillance beginning in childhood. Detailed management recommendations for individuals with PJS can be found in the NCCN Practice Guidelines and in the literature [173, 174].

PJS is caused by inherited mutations in the *STK11* gene (alias of *LKB1*) [175, 176]. It has been shown that when

both sequencing and deletion studies (via MLPA) of *STK11* are performed, 94 % of individuals who meet diagnostic criteria are found to have an *STK11* mutation [177]. The detection rate for those who meet these clinical criteria and have a positive family history approaches 100 %, whereas the detection rate is estimated to be 91 % when an individual meets clinical criteria but with absent family history [177]. Clinical genetic testing is available and can be used to help confirm a diagnosis of PJS as well as to provide predictive testing in asymptomatic individuals.

Amongst a series of 74 patients with SCTATs (the largest case series of this rare tumor), 27 (36.5 %) had a clinical diagnosis of Peutz-Jeghers syndrome [178]. The mean age of diagnosis in the PJS subgroup was 27 years compared to 34 years in the non-PJS subgroup. Differences in pathologic findings were noted between the subgroups of patients with and without PJS; SCTATs in patients with PJS were more frequently bilateral, multifocal, and with calcifications. Given that approximately one-third of SCTAT patients are likely to have PJS, it is reasonable to consider referring all SCTAT patients for genetic risk assessment. Identification of PJS in these patients prior to development of additional malignancies allows for targeted surveillance and prevention, and ideally, reduced morbidity and mortality.

Conclusion

Although each is rare in occurrence, these ten tumors are commonly associated with inherited cancer susceptibility. As summarized in Table 1, any patient with an adrenocortical carcinoma, thymic gland carcinoid tumor, fallopian tube cancer, primary peritoneal cancer, medullary thyroid cancer, paraganglioma, pheochromocytoma, sebaceous carcinoma, or sex cord tumor with annular tubules has a significant likelihood of carrying a germline mutation and warrants a thorough genetics work-up. Patients with diffuse gastric cancer at a young age, patients with bilateral or multifocal chromophobe, hybrid oncocytotic, and oncocytoma renal carcinomas, and patients with leiomyosarcoma and personal/family history suggestive of HLRCC or Lynch Syndrome should also be referred for a thorough genetics evaluation.

For individuals who are found to have one of the above cancer susceptibility syndromes, both the patient and his/her family are alerted to potential future cancer risks and can take advantage of appropriate surveillance and/or risk reduction. In general, prior to undergoing genetic testing, genetic risk assessment by a cancer genetics specialist (i.e. medical geneticist, genetic counselor, genetic clinical nurse, or other healthcare provider with genetics expertise) is recommended [79, 179, 180]. This allows for

identification of the appropriate gene/s and approach to genetic testing and it allows the patient to receive pre-test counseling, including a discussion of the risks, benefits, and limitations of genetic testing. In addition, genetic test results must be interpreted in the context of the personal and family history given current limitations in genetic testing to ensure appropriate medical management for the patient and the patient's family members.

Conflict of interest The authors declare that they have no conflict of interest.

References

- Bilimoria KY, Shen WT, Elaraj D, Bentrem DJ, Winchester DJ, Kebebew E, Sturgeon C (2008) Adrenocortical carcinoma in the United States: treatment utilization and prognostic factors. *Cancer* 113(11):3130–3136
- Wagner AS, Fleitz JM, Kleinschmidt-Demasters BK (2005) Pediatric adrenal cortical carcinoma: brain metastases and relationship to NF-1, case reports and review of the literature. *J Neurooncol* 75(2):127–133
- Skogseid B, Rastad J, Gobl A, Larsson C, Backlin K, Juhlin C, Akerstrom G, Oberg K (1995) Adrenal lesion in multiple endocrine neoplasia type 1. *Surgery* 118(6):1077–1082
- Seki M, Tanaka K, Kikuchi-Yanoshita R, Konishi M, Fukunari H, Iwama T, Miyaki M (1992) Loss of normal allele of the APC gene in an adrenocortical carcinoma from a patient with familial adenomatous polyposis. *Hum Genet* 89(3):298–300
- Sorensen SA, Mulvihill JJ, Nielsen A (1986) Long-term follow-up of von Recklinghausen neurofibromatosis. Survival and malignant neoplasms. *N Engl J Med* 314(16):1010–1015
- Marshall WH, Martin FI, Mackay IR (1967) Gardner's syndrome with adrenal carcinoma. *Australas Ann Med* 16(3):242–244
- Painter TA, Jagelman DG (1985) Adrenal adenomas and adrenal carcinomas in association with hereditary adenomatosis of the colon and rectum. *Cancer* 55(9):2001–2004
- Traill Z, Tuson J, Woodham C (1995) Adrenal carcinoma in a patient with Gardner's syndrome: imaging findings. *AJR Am J Roentgenol* 165(6):1460–1461
- Wakatsuki S, Sasano H, Matsui T, Nagashima K, Toyota T, Horii A (1998) Adrenocortical tumor in a patient with familial adenomatous polyposis: a case associated with a complete inactivating mutation of the APC gene and unusual histological features. *Hum Pathol* 29(3):302–306
- Lustbader ED, Williams WR, Bondy ML, Strom S, Strong LC (1992) Segregation analysis of cancer in families of childhood soft-tissue-sarcoma patients. *Am J Hum Genet* 51(2):344–356
- Hwang SJ, Lozano G, Amos CI, Strong LC (2003) Germline p53 mutations in a cohort with childhood sarcoma: sex differences in cancer risk. *Am J Hum Genet* 72(4):975–983
- Chompret A, Brugieres L, Ronsin M, Gardes M, Dessarps-Freichay F, Abel A, Hua D, Ligot L, Dondon MG, Bressac-de Paillerets B, Frebourg T, Lemerle J, Bonaiti-Pellie C, Feunteun J (2000) P53 germline mutations in childhood cancers and cancer risk for carrier individuals. *Br J Cancer* 82(12):1932–1937
- Hisada M, Garber JE, Fung CY, Fraumeni JF Jr, Li FP (1998) Multiple primary cancers in families with Li-Fraumeni syndrome. *J Natl Cancer Inst* 90(8):606–611
- Garber JE, Goldstein AM, Kantor AF, Dreyfus MG, Fraumeni JF Jr, Li FP (1991) Follow-up study of twenty-four families with Li-Fraumeni syndrome. *Cancer Res* 51(22):6094–6097
- Nichols KE, Malkin D, Garber JE, Fraumeni JF Jr, Li FP (2001) Germ-line p53 mutations predispose to a wide spectrum of early-onset cancers. *Cancer Epidemiol Biomarkers Prev* 10(2):83–87
- Olivier M, Goldgar DE, Sodha N, Ohgaki H, Kleihues P, Hainaut P, Eeles RA (2003) Li-Fraumeni and related syndromes: correlation between tumor type, family structure, and TP53 genotype. *Cancer Res* 63(20):6643–6650
- Li FP, Fraumeni JF Jr, Mulvihill JJ, Blattner WA, Dreyfus MG, Tucker MA, Miller RW (1988) A cancer family syndrome in twenty-four kindreds. *Cancer Res* 48(18):5358–5362
- Brugieres L, Gardes M, Moutou C, Chompret A, Meresse V, Martin A, Poisson N, Flamant F, Bonaiti-Pellie C, Lemerle J et al (1993) Screening for germ line p53 mutations in children with malignant tumors and a family history of cancer. *Cancer Res* 53(3):452–455
- Frebourg T, Barbier N, Yan YX, Garber JE, Dreyfus M, Fraumeni J Jr, Li FP, Friend SH (1995) Germ-line p53 mutations in 15 families with Li-Fraumeni syndrome. *Am J Hum Genet* 56(3):608–615
- Birch JM, Hartley AL, Tricker KJ, Prosser J, Condie A, Kelsey AM, Harris M, Jones PH, Binchy A, Crowther D et al (1994) Prevalence and diversity of constitutional mutations in the p53 gene among 21 Li-Fraumeni families. *Cancer Res* 54(5):1298–1304
- Birch JM, Heighway J, Teare MD, Kelsey AM, Hartley AL, Tricker KJ, Crowther D, Lane DP, Santibanez-Koref MF (1994) Linkage studies in a Li-Fraumeni family with increased expression of p53 protein but no germline mutation in p53. *Br J Cancer* 70(6):1176–1181
- Varley JM, McGown G, Thorncroft M, Santibanez-Koref MF, Kelsey AM, Tricker KJ, Evans DG, Birch JM (1997) Germ-line mutations of TP53 in Li-Fraumeni families: an extended study of 39 families. *Cancer Res* 57(15):3245–3252
- Chompret A, Abel A, Stoppa-Lyonnet D, Brugieres L, Pages S, Feunteun J, Bonaiti-Pellie C (2001) Sensitivity and predictive value of criteria for p53 germline mutation screening. *J Med Genet* 38(1):43–47
- Gonzalez KD, Buzin CH, Noltner KA, Gu D, Li W, Malkin D, Sommer SS (2009) High frequency of de novo mutations in Li-Fraumeni syndrome. *J Med Genet* 46(10):689–693
- Gonzalez KD, Noltner KA, Buzin CH, Gu D, Wen-Fong CY, Nguyen VQ, Han JH, Lowstuter K, Longmate J, Sommer SS, Weitzel JN (2009) Beyond Li Fraumeni Syndrome: clinical characteristics of families with p53 germline mutations. *J Clin Oncol* 27(8):1250–1256
- Tinat J, Bougeard G, Baert-Desurmont S, Vasseur S, Martin C, Bouvignies E, Caron O, Bressac-de Paillerets B, Berthet P, Dugast C, Bonaiti-Pellie C, Stoppa-Lyonnet D, Frebourg T (2009) 2009 version of the Chompret criteria for Li Fraumeni syndrome. *J Clin Oncol* 27(26):e108–e109; author reply e110
- Villani A, Tabori U, Schiffman J, Shlien A, Beyene J, Druker H, Novokmet A, Finlay J, Malkin D (2011) Biochemical and imaging surveillance in germline TP53 mutation carriers with Li-Fraumeni syndrome: a prospective observational study. *Lancet Oncol* 12(6):559–567. doi:10.1016/S1470-2045(11)70119-X
- Lapunzina P (2005) Risk of tumorigenesis in overgrowth syndromes: a comprehensive review. *Am J Med Genet C Semin Med Genet* 137C(1):53–71
- Cohen MM Jr (2005) Beckwith-Wiedemann syndrome: historical, clinicopathological, and etiopathogenetic perspectives. *Pediatr Dev Pathol* 8(3):287–304
- Martinez Y, Martinez R (1996) Clinical features in the Wiedemann-Beckwith syndrome. *Clin Genet* 50(4):272–274
- Modlin IM, Sandor A (1997) An analysis of 8305 cases of carcinoid tumors. *Cancer* 79(4):813–829
- Godwin JD 2nd (1975) Carcinoid tumors. An analysis of 2,837 cases. *Cancer* 36(2):560–569

33. Rodriguez-Bigas MA, Vasen HF, Lynch HT, Watson P, Myrhoj T, Jarvinen HJ, Mecklin JP, Macrae F, St John DJ, Bertario L, Fidalgo P, Madlensky L, Rozen P (1998) Characteristics of small bowel carcinoma in hereditary nonpolyposis colorectal carcinoma. International Collaborative Group on HNPCC. *Cancer* 83(2):240–244
34. Marx S, Spiegel AM, Skarulis MC, Doppman JL, Collins FS, Liotta LA (1998) Multiple endocrine neoplasia type 1: clinical and genetic topics. *Ann Intern Med* 129(6):484–494
35. Cappelli C, Agosti B, Braga M, Cumetti D, Gandossi E, Rizzoni D, Agabiti Rosei E (2004) Von Recklinghausen's neurofibromatosis associated with duodenal somatostatinoma. A case report and review of the literature. *Minerva Endocrinol* 29(1):19–24
36. Sachithanandan N, Harle RA, Burgess JR (2005) Bronchopulmonary carcinoid in multiple endocrine neoplasia type 1. *Cancer* 103(3):509–515
37. Teh BT, McArdle J, Chan SP, Menon J, Hartley L, Pullan P, Ho J, Khir A, Wilkinson S, Larsson C, Cameron D, Shepherd J (1997) Clinicopathologic studies of thymic carcinoids in multiple endocrine neoplasia type 1. *Medicine (Baltimore)* 76(1):21–29
38. Debelenko LV, Emmert-Buck MR, Zhuang Z, Epshteyn E, Moskaluk CA, Jensen RT, Liotta LA, Lubensky IA (1997) The multiple endocrine neoplasia type I gene locus is involved in the pathogenesis of type II gastric carcinoids. *Gastroenterology* 113(3):773–781
39. Bassett JH, Forbes SA, Pannett AA, Lloyd SE, Christie PT, Wooding C, Harding B, Besser GM, Edwards CR, Monson JP, Sampson J, Wass JA, Wheeler MH, Thakker RV (1998) Characterization of mutations in patients with multiple endocrine neoplasia type 1. *Am J Hum Genet* 62(2):232–244
40. Marx SJ (2001) Multiple endocrine neoplasia type 1. In: Scriver CR, Beaudet AL, Sly WS, Valle D (eds) *The metabolic and molecular bases of inherited disease*, 8th edn. McGraw-Hill, New York, pp 943–966
41. Brandi ML, Gagel RF, Angeli A, Bilezikian JP, Beck-Peccoz P, Bordi C, Conte-Devolx B, Falchetti A, Gheri RG, Libroia A, Lips CJ, Lombardi G, Mannelli M, Pacini F, Ponder BA, Raue F, Skogseid B, Tamburrano G, Thakker RV, Thompson NW, Tomassetti P, Tonelli F, Wells SA Jr, Marx SJ (2001) Guidelines for diagnosis and therapy of MEN type 1 and type 2. *J Clin Endocrinol Metab* 86(12):5658–5671
42. Bordi C, Falchetti A, Azzoni C, D'Adda T, Canavese G, Guariglia A, Santini D, Tomassetti P, Brandi ML (1997) Aggressive forms of gastric neuroendocrine tumors in multiple endocrine neoplasia type I. *Am J Surg Pathol* 21(9):1075–1082
43. Jensen RT (1999) MEN-1 carcinoids: diagnosis and therapy. In: *Proceedings of the 7th international workshop on multiple neoplasia*, Gubbio, Italy
44. WHO (2011) Fact sheet no 297: Cancer. <http://www.who.int/mediacentre/factsheets/fs297/en/>
45. Fitzgerald RC, Hardwick R, Huntsman D, Carneiro F, Guilford P, Blair V, Chung DC, Norton J, Ragunath K, Van Krieken JH, Dwerryhouse S, Caldas C (2010) Hereditary diffuse gastric cancer: updated consensus guidelines for clinical management and directions for future research. *J Med Genet* 47(7):436–444
46. Campbell F, Lauwers GY, Williams GT (2007) In: Fletcher CDM (ed) *Diagnostic Histopathology of Tumors*, vol 1, 3rd edn. Churchill Livingstone Elsevier, pp 327–378
47. Crew KD, Neugut AI (2006) Epidemiology of gastric cancer. *World J Gastroenterol* 12(3):354–362
48. Caldas C, Carneiro F, Lynch HT, Yokota J, Wiesner GL, Powell SM, Lewis FR, Huntsman DG, Pharoah PD, Jankowski JA, MacLeod P, Vogelsang H, Keller G, Park KG, Richards FM, Maher ER, Gayther SA, Oliveira C, Grehan N, Wight D, Seruca R, Roviello F, Ponder BA, Jackson CE (1999) Familial gastric cancer: overview and guidelines for management. *J Med Genet* 36(12):873–880
49. Oliveira C, Bordin MC, Grehan N, Huntsman D, Suriano G, Machado JC, Kiviluoto T, Aaltonen L, Jackson CE, Seruca R, Caldas C (2002) Screening E-cadherin in gastric cancer families reveals germline mutations only in hereditary diffuse gastric cancer kindred. *Hum Mutat* 19(5):510–517
50. Park JG, Yang HK, Kim WH, Caldas C, Yokota J, Guilford PJ (2000) Report on the first meeting of the International Collaborative Group on Hereditary Gastric Cancer. *J Natl Cancer Inst* 92(21):1781–1782
51. Lynch HT, Grady W, Lynch JF, Tsuchiya KD, Wiesner G, Markowitz SD (2000) E-cadherin mutation-based genetic counseling and hereditary diffuse gastric carcinoma. *Cancer Genet Cytogenet* 122(1):1–6
52. Oliveira C, Seruca R, Carneiro F (2006) Genetics, pathology, and clinics of familial gastric cancer. *Int J Surg Pathol* 14(1):21–33
53. Kaurah P, MacMillan A, Boyd N, Senz J, De Luca A, Chun N, Suriano G, Zaor S, Van Manen L, Gilpin C, Nikkel S, Connolly-Wilson M, Weissman S, Rubinstein WS, Sebold C, Greenstein R, Stroop J, Yim D, Panzini B, McKinnon W, Greenblatt M, Wirtzfeld D, Fontaine D, Coit D, Yoon S, Chung D, Lauwers G, Pizzuti A, Vaccaro C, Redal MA, Oliveira C, Tischkowitz M, Olschwang S, Gallinger S, Lynch H, Green J, Ford J, Pharoah P, Fernandez B, Huntsman D (2007) Founder and recurrent CDH1 mutations in families with hereditary diffuse gastric cancer. *JAMA* 297(21):2360–2372
54. Oliveira C, Sousa S, Pinheiro H, Karam R, Bordeira-Carrico R, Senz J, Kaurah P, Carvalho J, Pereira R, Gusmao L, Wen X, Cipriano MA, Yokota J, Carneiro F, Huntsman D, Seruca R (2009) Quantification of epigenetic and genetic 2nd hits in CDH1 during hereditary diffuse gastric cancer syndrome progression. *Gastroenterology* 136(7):2137–2148
55. Pharoah PD, Guilford P, Caldas C (2001) Incidence of gastric cancer and breast cancer in CDH1 (E-cadherin) mutation carriers from hereditary diffuse gastric cancer families. *Gastroenterology* 121(6):1348–1353
56. Chun YS, Lindor NM, Smyrk TC, Petersen BT, Burgart LJ, Guilford PJ, Donohue JH (2001) Germline E-cadherin gene mutations: is prophylactic total gastrectomy indicated? *Cancer* 92(1):181–187
57. Huntsman DG, Carneiro F, Lewis FR, MacLeod PM, Hayashi A, Monaghan KG, Maung R, Seruca R, Jackson CE, Caldas C (2001) Early gastric cancer in young, asymptomatic carriers of germ-line E-cadherin mutations. *N Engl J Med* 344(25):1904–1909
58. Suriano G, Yew S, Ferreira P, Senz J, Kaurah P, Ford JM, Longacre TA, Norton JA, Chun N, Young S, Oliveira MJ, Macgillivray B, Rao A, Sears D, Jackson CE, Boyd J, Yee C, Deters C, Pai GS, Hammond LS, McGivern BJ, Medgyesy D, Sartz D, Arun B, Oelschlagel BK, Upton MP, Neufeld-Kaiser W, Silva OE, Donenberg TR, Kooby DA, Sharma S, Jonsson BA, Gronberg H, Gallinger S, Seruca R, Lynch H, Huntsman DG (2005) Characterization of a recurrent germ line mutation of the E-cadherin gene: implications for genetic testing and clinical management. *Clin Cancer Res* 11(15):5401–5409
59. Roffers SD, Wu XC, Johnson CH, Correa CN (2003) Incidence of extraovarian primary cancers in the United States, 1992–1997. *Cancer* 97(10 Suppl):2643–2647
60. Levine DA, Argenta PA, Yee CJ, Marshall DS, Olvera N, Bogomolny F, Rahaman JA, Robson ME, Offit K, Barakat RR, Soslow RA, Boyd J (2003) Fallopian tube and primary peritoneal carcinomas associated with BRCA mutations. *J Clin Oncol* 21(22):4222–4227
61. Zweemer RP, van Diest PJ, Verheijen RH, Ryan A, Gille JJ, Sijmons RH, Jacobs IJ, Menko FH, Kenemans P (2000) Molecular evidence linking primary cancer of the fallopian tube to BRCA1 germline mutations. *Gynecol Oncol* 76(1):45–50

62. Sobol H, Jacquemier J, Bonaiti C, Dauplat J, Birnbaum D, Eisinger F (2000) Fallopian tube cancer as a feature of BRCA1-associated syndromes. *Gynecol Oncol* 78(2):263–264
63. Antoniou A, Pharoah PD, Narod S, Risch HA, Eyfjord JE, Hopper JL, Loman N, Olsson H, Johannsson O, Borg A, Pasini B, Radice P, Manoukian S, Eccles DM, Tang N, Olah E, Anton-Culver H, Warner E, Lubinski J, Gronwald J, Gorski B, Tulinius H, Thorlacius S, Eerola H, Nevanlinna H, Syrjakoski K, Kallioniemi OP, Thompson D, Evans C, Peto J, Lalloo F, Evans DG, Easton DF (2003) Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case Series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet* 72(5):1117–1130
64. Chen S, Parmigiani G (2007) Meta-analysis of BRCA1 and BRCA2 penetrance. *J Clin Oncol* 25(11):1329–1333
65. Ford D, Easton DF, Bishop DT, Narod SA, Goldgar DE (1994) Risks of cancer in BRCA1-mutation carriers. *Breast Cancer Linkage Consortium. Lancet* 343(8899):692–695
66. Ford D, Easton DF, Stratton M, Narod S, Goldgar D, Devilee P, Bishop DT, Weber B, Lenoir G, Chang-Claude J, Sobol H, Teare MD, Struwing J, Arason A, Scherneck S, Peto J, Rebbeck TR, Tonin P, Neuhausen S, Barkardottir R, Eyfjord J, Lynch H, Ponder BA, Gayther SA, Zelada-Hedman M et al (1998) Genetic heterogeneity and penetrance analysis of the BRCA1 and BRCA2 genes in breast cancer families. *The Breast Cancer Linkage Consortium. Am J Hum Genet* 62(3):676–689
67. King MC, Marks JH, Mandell JB (2003) Breast and ovarian cancer risks due to inherited mutations in BRCA1 and BRCA2. *Science* 302(5645):643–646
68. SEER Cancer Statistics Review, 1975–2005 (2008) Surveillance Epidemiology and End Results (SEER). www.seer.cancer.gov/csr/1975_2005/
69. Finch A, Beiner M, Lubinski J, Lynch HT, Moller P, Rosen B, Murphy J, Ghadirian P, Friedman E, Foulkes WD, Kim-Sing C, Wagner T, Tung N, Couch F, Stoppa-Lyonnet D, Ainsworth P, Daly M, Pasini B, Gershoni-Baruch R, Eng C, Olopade OI, McLennan J, Karlan B, Weitzel J, Sun P, Narod SA (2006) Salpingo-oophorectomy and the risk of ovarian, fallopian tube, and peritoneal cancers in women with a BRCA1 or BRCA2 Mutation. *JAMA* 296(2):185–192
70. Thompson D, Easton DF (2002) Cancer incidence in BRCA1 mutation carriers. *J Natl Cancer Inst* 94(18):1358–1365
71. Tai YC, Domchek S, Parmigiani G, Chen S (2007) Breast cancer risk among male BRCA1 and BRCA2 mutation carriers. *J Natl Cancer Inst* 99(23):1811–1814
72. Cancer Linkage Consortium (1999) Cancer risks in BRCA2 mutation carriers. *The Breast Cancer Linkage Consortium. J Natl Cancer Inst* 91(15):1310–1316
73. Menczer J, Chetrit A, Barda G, Lubin F, Fishler Y, Altaras M, Levavi H, Struwing JP, Sadetzki S, Modan B (2003) Frequency of BRCA mutations in primary peritoneal carcinoma in Israeli Jewish women. *Gynecol Oncol* 88(1):58–61
74. Aziz S, Kuperstein G, Rosen B, Cole D, Nedelcu R, McLaughlin J, Narod SA (2001) A genetic epidemiological study of carcinoma of the fallopian tube. *Gynecol Oncol* 80(3):341–345
75. Cass I, Holschneider C, Datta N, Barbuto D, Walts AE, Karlan BY (2005) BRCA-mutation-associated fallopian tube carcinoma: a distinct clinical phenotype? *Obstet Gynecol* 106(6):1327–1334
76. Lu K, Kauff N, Powell CB, Chen LM, Cass I, Lancaster J, Karlan B, Berchuck A, Mutch D (2009) ACOG Practice Bulletin No. 103: hereditary breast and ovarian cancer syndrome. *Obstet Gynecol* 113(4):957–966
77. Powell CB, Kenley E, Chen LM, Crawford B, McLennan J, Zaloudek C, Komaromy M, Beattie M, Ziegler J (2005) Risk-reducing salpingo-oophorectomy in BRCA mutation carriers: role of serial sectioning in the detection of occult malignancy. *J Clin Oncol* 23(1):127–132
78. Rabban JT, Krasik E, Chen LM, Powell CB, Crawford B, Zaloudek CJ (2009) Multistep level sections to detect occult fallopian tube carcinoma in risk-reducing salpingo-oophorectomies from women with BRCA mutations: implications for defining an optimal specimen dissection protocol. *Am J Surg Pathol* 33(12):1878–1885
79. Daly MB, Axilbund JE, Buys S, Crawford B, Farrell CD, Friedman S, Garber JE, Goorha S, Gruber SB, Hampel H, Kakkiani V, Kohlmann W, Kurian A, Litton J, Marcom PK, Nussbaum R, Offit K, Pal T, Pasche B, Pilarski R, Reiser G, Shannon KM, Smith JR, Swisher E, Weitzel JN (2010) Genetic/familial high-risk assessment: breast and ovarian. *J Natl Compr Canc Netw* 8(5):562–594
80. Toro JR, Travis LB, Wu HJ, Zhu K, Fletcher CD, Devesa SS (2006) Incidence patterns of soft tissue sarcomas, regardless of primary site, in the surveillance, epidemiology and end results program, 1978–2001: an analysis of 26,758 cases. *Int J Cancer* 119(12):2922–2930
81. Enzinger FM, Weiss SW (1995) Leiomyosarcoma. In: Gay SM (ed) *Soft tissue tumors*, 3rd edn. Mosby-Year Book, St. Louis, pp 491–510
82. Blanco A, Grana B, Fachal L, Santamarina M, Cameselle-Teijeiro J, Ruiz-Ponte C, Carracedo A, Vega A (2010) Beyond BRCA1 and BRCA2 wild-type breast and/or ovarian cancer families: germline mutations in TP53 and PTEN. *Clin Genet* 77(2):193–196
83. Varley JM (2003) Germline TP53 mutations and Li-Fraumeni syndrome. *Hum Mutat* 21(3):313–320
84. Manoukian S, Peissel B, Pensotti V, Barile M, Cortesi L, Stacchiotti S, Terenziani M, Barbera F, Pasquini G, Frigerio S, Pierotti MA, Radice P, Della-Torre G (2007) Germline mutations of TP53 and BRCA2 genes in breast cancer/sarcoma families. *Eur J Cancer* 43(3):601–606
85. Launonen V, Vierimaa O, Kiuru M, Isola J, Roth S, Pukkala E, Sistonen P, Herva R, Aaltonen LA (2001) Inherited susceptibility to uterine leiomyomas and renal cell cancer. *Proc Natl Acad Sci USA* 98(6):3387–3392
86. Toro JR, Nickerson ML, Wei MH, Warren MB, Glenn GM, Turner ML, Stewart L, Duray P, Tourre O, Sharma N, Choyke P, Stratton P, Merino M, Walther MM, Linehan WM, Schmidt LS, Zbar B (2003) Mutations in the fumarate hydratase gene cause hereditary leiomyomatosis and renal cell cancer in families in North America. *Am J Hum Genet* 73(1):95–106
87. Alam NA, Rowan AJ, Wortham NC, Pollard PJ, Mitchell M, Tyrer JP, Barclay E, Calonje E, Manek S, Adams SJ, Bowers PW, Burrows NP, Charles-Holmes R, Cook LJ, Daly BM, Ford GP, Fuller LC, Hadfield-Jones SE, Hardwick N, Highet AS, Keefe M, MacDonald-Hull SP, Potts ED, Crone M, Wilkinson S, Camacho-Martinez F, Jablonska S, Ratnavel R, MacDonald A, Mann RJ, Grice K, Guillet G, Lewis-Jones MS, McGrath H, Seukeran DC, Morrison PJ, Fleming S, Rahman S, Kelsell D, Leigh I, Olpin S, Tomlinson IP (2003) Genetic and functional analyses of FH mutations in multiple cutaneous and uterine leiomyomatosis, hereditary leiomyomatosis and renal cancer, and fumarate hydratase deficiency. *Hum Mol Genet* 12(11):1241–1252
88. Alam NA, Barclay E, Rowan AJ, Tyrer JP, Calonje E, Manek S, Kelsell D, Leigh I, Olpin S, Tomlinson IP (2005) Clinical features of multiple cutaneous and uterine leiomyomatosis: an underdiagnosed tumor syndrome. *Arch Dermatol* 141(2):199–206
89. Lehtonen HJ, Kiuru M, Ylisaukko-Oja SK, Salovaara R, Herva R, Koivisto PA, Vierimaa O, Aittomaki K, Pukkala E, Launonen V, Aaltonen LA (2006) Increased risk of cancer in patients with

- fumarate hydratase germline mutation. *J Med Genet* 43(6): 523–526
90. Stewart L, Glenn GM, Stratton P, Goldstein AM, Merino MJ, Tucker MA, Linehan WM, Toro JR (2008) Association of germline mutations in the fumarate hydratase gene and uterine fibroids in women with hereditary leiomyomatosis and renal cell cancer. *Arch Dermatol* 144(12):1584–1592
 91. Schneider K (2011) Counseling about cancer: strategies for Genetic Counseling, 3rd edn
 92. Smit DL, Mensenkamp AR, Badeloe S, Breuning MH, Simon ME, van Spaendonck KY, Aalfs CM, Post JG, Shanley S, Krapels IP, Hoefsloot LH, van Moorselaar RJ, Starink TM, Bayley JP, Frank J, van Steensel MA, Menko FH (2010) Hereditary leiomyomatosis and renal cell cancer in families referred for fumarate hydratase germline mutation analysis. *Clin Genet* 79(1):49–59
 93. Refae MA, Wong N, Patenaude F, Begin LR, Foulkes WD (2007) Hereditary leiomyomatosis and renal cell cancer: an unusual and aggressive form of hereditary renal carcinoma. *Nat Clin Pract* 4(4):256–261
 94. Lynch HT, Smyrk T (1996) Hereditary nonpolyposis colorectal cancer (Lynch syndrome). An updated review. *Cancer* 78(6): 1149–1167
 95. Vasen HF, Wijnen JT, Menko FH, Kleibeuker JH, Taal BG, Griffioen G, Nagengast FM, Meijers-Heijboer EH, Bertario L, Varesco L, Bisgaard ML, Mohr J, Fodde R, Khan PM (1996) Cancer risk in families with hereditary nonpolyposis colorectal cancer diagnosed by mutation analysis. *Gastroenterology* 110(4):1020–1027
 96. Lin KM, Shashidharan M, Thorson AG, Terment CA, Blatchford GJ, Christensen MA, Watson P, Lemon SJ, Franklin B, Karr B, Lynch J, Lynch HT (1998) Cumulative incidence of colorectal and extracolonic cancers in MLH1 and MSH2 mutation carriers of hereditary nonpolyposis colorectal cancer. *J Gastrointest Surg* 2(1):67–71
 97. Peltomaki P (2003) Role of DNA mismatch repair defects in the pathogenesis of human cancer. *J Clin Oncol* 21(6):1174–1179
 98. Kovacs ME, Papp J, Szentirmay Z, Otto S, Olah E (2009) Deletions removing the last exon of TACSTD1 constitute a distinct class of mutations predisposing to Lynch syndrome. *Hum Mutat* 30(2):197–203. doi:10.1002/humu.20942
 99. Medina Arana V, Barrios del Pino Y, Garcia-Castro C, Gonzalez-Aguilera JJ, Fernandez-Peralta A, Gonzalez Hermoso F (2002) Highly aggressive leiomyosarcoma associated with Lynch II syndrome: increasing the range of extracolonic cancers related with hereditary non-polyposis colonic cancer. *Ann Oncol* 13(5):807–808
 100. Nilbert M, Therkildsen C, Nissen A, Akerman M, Bernstein I (2009) Sarcomas associated with hereditary nonpolyposis colorectal cancer: broad anatomical and morphological spectrum. *Fam Cancer* 8(3):209–213
 101. Kruse R, Rutten A, Lamberti C, Hosseiny-Malayeri HR, Wang Y, Ruelfs C, Jungck M, Mathiak M, Ruzicka T, Hartschuh W, Bisceglia M, Friedl W, Propping P (1998) Muir-Torre phenotype has a frequency of DNA mismatch-repair-gene mutations similar to that in hereditary nonpolyposis colorectal cancer families defined by the Amsterdam criteria. *Am J Hum Genet* 63(1):63–70
 102. Venkatraman L, Goepel JR, Steele K, Dobbs SP, Lyness RW, McCluggage WG (2003) Soft tissue, pelvic, and urinary bladder leiomyosarcoma as second neoplasm following hereditary retinoblastoma. *J Clin Pathol* 56(3):233–236
 103. Kleiner RA, Tucker MA, Abramson DH, Seddon JM, Tarone RE, Fraumeni JF Jr (2007) Risk of soft tissue sarcomas by individual subtype in survivors of hereditary retinoblastoma. *J Natl Cancer Inst* 99(1):24–31
 104. Brucker B, Ernst L, Meadows A, Zderic S (2006) A second leiomyosarcoma in the urinary bladder of a child with a history of retinoblastoma 12 years following partial cystectomy. *Pediatr Blood Cancer* 46(7):811–814
 105. Moley JF (1997) Medullary thyroid carcinoma. In: Clark OH, Duh QY (eds) *Textbook of endocrine surgery*. W.B. Saunders, Philadelphia, pp 108–118
 106. Kloos RT, Eng C, Evans DB, Francis GL, Gagel RF, Gharib H, Moley JF, Pacini F, Ringel MD, Schlumberger M, Wells SA Jr (2009) Medullary thyroid cancer: management guidelines of the American Thyroid Association. *Thyroid* 19(6):565–612
 107. Morrison PJ, Nevin NC (1996) Multiple endocrine neoplasia type 2B (mucosal neuroma syndrome, Wagenmann-Froboese syndrome). *J Med Genet* 33(9):779–782
 108. Eng C, Clayton D, Schuffenecker I, Lenoir G, Cote G, Gagel RF, van Amstel HK, Lips CJ, Nishisho I, Takai SI, Marsh DJ, Robinson BG, Frank-Raue K, Raue F, Xue F, Noll WW, Romei C, Pacini F, Fink M, Niederle B, Zedenius J, Nordenskjold M, Komminoth P, Hendy GN, Mulligan LM et al (1996) The relationship between specific RET proto-oncogene mutations and disease phenotype in multiple endocrine neoplasia type 2. International RET mutation consortium analysis. *JAMA* 276(19):1575–1579
 109. Elisei R, Romei C, Cosci B, Agate L, Bottici V, Molinaro E, Sculli M, Miccoli P, Basolo F, Grasso L, Pacini F, Pinchera A (2007) RET genetic screening in patients with medullary thyroid cancer and their relatives: experience with 807 individuals at one center. *J Clin Endocrinol Metab* 92(12):4725–4729
 110. Cohen MS, Moley JF (2003) Surgical treatment of medullary thyroid carcinoma. *J Intern Med* 253(6):616–626
 111. Kimura N, Chetty R, Capella C, Young WF Jr, Koch CA, Lam KY, DeLellis RA, Kawashima A, Komminoth P, Tischler AS (2004) Pathology and genetics: tumours of the endocrine organs. In: DeLellis RA, Lloyd RV, Heitz PU, Eng C (eds) *World Health Organization classification of tumours*. Oxford University Press, Oxford, p 159
 112. Benn DE, Gimenez-Roqueplo AP, Reilly JR, Bertherat J, Burgess J, Byth K, Croxson M, Dahia PL, Elston M, Gimm O, Henley D, Herman P, Murday V, Niccoli-Sire P, Pasiaka JL, Rohrer V, Tucker K, Jeunemaitre X, Marsh DJ, Plouin PF, Robinson BG (2006) Clinical presentation and penetrance of pheochromocytoma/paraganglioma syndromes. *J Clin Endocrinol Metab* 91(3):827–836
 113. Eric Z, Neumann HP (2009) When should genetic testing be obtained in a patient with pheochromocytoma or paraganglioma? *Clin Endocrinol (Oxf)* 70(3):354–357
 114. Bayley JP, Kunst HP, Cascon A, Sampietro ML, Gaal J, Korpershoek E, Hinojar-Gutierrez A, Timmers HJ, Hoefsloot LH, Hermens MA, Suarez C, Hussain AK, Vriends AH, Hes FJ, Jansen JC, Tops CM, Corssmit EP, de Knijff P, Lenders JW, Cremers CW, Devilee P, Dinjens WN, de Krijger RR, Robledo M (2010) SDHAF2 mutations in familial and sporadic paraganglioma and pheochromocytoma. *Lancet Oncol* 11(4):366–372
 115. Qin Y, Yao L, King EE, Buddavarapu K, Lenci RE, Chocron ES, Lechleiter JD, Sass M, Aronin N, Schiavi F, Boaretto F, Opocher G, Toledo RA, Toledo SP, Stiles C, Aguiar RC, Dahia PL (2010) Germline mutations in TMEM127 confer susceptibility to pheochromocytoma. *Nat Genet* 42(3):229–233
 116. Burnichon N, Briere JJ, Libe R, Vescovo L, Riviere J, Tissier F, Jouanno E, Jeunemaitre X, Benit P, Tzagoloff A, Rustin P, Bertherat J, Favier J, Gimenez-Roqueplo AP (2010) SDHA is a tumor suppressor gene causing paraganglioma. *Hum Mol Genet* 19(15):3011–3020. doi:10.1093/hmg/ddq206
 117. Comino-Mendez I, Gracia-Aznarez FJ, Schiavi F, Landa I, Leandro-Garcia LJ, Leton R, Honrado E, Ramos-Medina R,

- Caronia D, Pita G, Gomez-Grana A, de Cubas AA, Inglada-Perez L, Maliszewska A, Taschin E, Bobisse S, Pica G, Loli P, Hernandez-Lavado R, Diaz JA, Gomez-Morales M, Gonzalez-Neira A, Roncador G, Rodriguez-Antona C, Benitez J, Mannelli M, Opocher G, Robledo M, Cascon A (2011) Exome sequencing identifies MAX mutations as a cause of hereditary pheochromocytoma. *Nat Genet* 43(7):663–667. doi:[10.1038/ng.861](https://doi.org/10.1038/ng.861)
118. Rodriguez JM, Balsalobre M, Ponce JL, Rios A, Torregrosa NM, Tebar J, Parrilla P (2008) Pheochromocytoma in MEN 2A syndrome. Study of 54 patients. *World J Surg* 32(11):2520–2526
119. Inabnet WB, Caragliano P, Pertsemlidis D (2000) Pheochromocytoma: inherited associations, bilaterality, and cortex preservation. *Surgery* 128(6):1007–1011;discussion 1011–1002
120. Pacak K, Ilias I, Adams KT, Eisenhofer G (2005) Biochemical diagnosis, localization and management of pheochromocytoma: focus on multiple endocrine neoplasia type 2 in relation to other hereditary syndromes and sporadic forms of the tumour. *J Intern Med* 257(1):60–68
121. Pomares FJ, Canas R, Rodriguez JM, Hernandez AM, Parrilla P, Tebar FJ (1998) Differences between sporadic and multiple endocrine neoplasia type 2A pheochromocytoma. *Clin Endocrinol (Oxf)* 48(2):195–200
122. Neumann HP, Eng C (2009) The approach to the patient with paraganglioma. *J Clin Endocrinol Metab* 94(8):2677–2683
123. Maher ER, Webster AR, Richards FM, Green JS, Crossey PA, Payne SJ, Moore AT (1996) Phenotypic expression in von Hippel-Lindau disease: correlations with germline VHL gene mutations. *J Med Genet* 33(4):328–332
124. Hao HX, Khalimonchuk O, Schraders M, Dephore N, Bayley JP, Kunst H, Devilee P, Cremers CW, Schiffman JD, Bentz BG, Gygi SP, Winge DR, Kremer H, Rutter J (2009) SDH5, a gene required for flavination of succinate dehydrogenase, is mutated in paraganglioma. *Science* 325(5944):1139–1142
125. Neumann HP, Pawlu C, Peczkowska M, Bausch B, McWhinney SR, Muresan M, Buchta M, Franke G, Klisch J, Bley TA, Hoegerle S, Boedeker CC, Opocher G, Schipper J, Januszewicz A, Eng C (2004) Distinct clinical features of paraganglioma syndromes associated with SDHB and SDHD gene mutations. *JAMA* 292(8):943–951
126. Schiavi F, Boedeker CC, Bausch B, Peczkowska M, Gomez CF, Strassburg T, Pawlu C, Buchta M, Salzmann M, Hoffmann MM, Berlis A, Brink I, Cybulla M, Muresan M, Walter MA, Forrer F, Valimaki M, Kawecky A, Szutkowski Z, Schipper J, Walz MK, Pigny P, Bauters C, Willet-Brozick JE, Baysal BE, Januszewicz A, Eng C, Opocher G, Neumann HP (2005) Predictors and prevalence of paraganglioma syndrome associated with mutations of the SDHC gene. *JAMA* 294(16):2057–2063
127. Pasini B, McWhinney SR, Bei T, Matyakhina L, Stergiopoulos S, Muchow M, Boikos SA, Ferrando B, Pacak K, Assie G, Baudin E, Chompret A, Ellison JW, Briere JJ, Rustin P, Gimenez-Roqueplo AP, Eng C, Carney JA, Stratakis CA (2008) Clinical and molecular genetics of patients with the Carney-Stratakis syndrome and germline mutations of the genes coding for the succinate dehydrogenase subunits SDHB, SDHC, and SDHD. *Eur J Hum Genet* 16(1):79–88
128. Vanharanta S, Buchta M, McWhinney SR, Virta SK, Peczkowska M, Morrison CD, Lehtonen R, Januszewicz A, Jarvinen H, Juhola M, Mecklin JP, Pukkala E, Herva R, Kiuru M, Nupponen NN, Aaltonen LA, Neumann HP, Eng C (2004) Early-onset renal cell carcinoma as a novel extraparaganglial component of SDHB-associated heritable paraganglioma. *Am J Hum Genet* 74(1):153–159
129. Ricketts C, Woodward ER, Killick P, Morris MR, Astuti D, Latif F, Maher ER (2008) Germline SDHB mutations and familial renal cell carcinoma. *J Natl Cancer Inst* 100(17):1260–1262
130. Boedeker CC, Neumann HP, Offergeld C, Maier W, Falcioni M, Berlis A, Schipper J (2009) Clinical features of paraganglioma syndromes. *Skull Base* 19(1):17–25
131. Bausch B, Borozdin W, Mautner VF, Hoffmann MM, Boehm D, Robledo M, Cascon A, Harenberg T, Schiavi F, Pawlu C, Peczkowska M, Letizia C, Calvieri S, Arnaldi G, Klingenberg-Noftz RD, Reisch N, Fassina A, Brunaud L, Walter MA, Mannelli M, MacGregor G, Palazzo FF, Barontini M, Walz MK, Kremens B, Brabant G, Pfaffle R, Koschker AC, Lohofner F, Mohaupt M, Gimm O, Jarzab B, McWhinney SR, Opocher G, Januszewicz A, Kohlhase J, Eng C, Neumann HP (2007) Germline NF1 mutational spectra and loss-of-heterozygosity analyses in patients with pheochromocytoma and neurofibromatosis type 1. *J Clin Endocrinol Metab* 92(7):2784–2792
132. (1988) National Institutes of Health Consensus Development Conference Statement: neurofibromatosis. Bethesda, Md., USA, July 13–15, 1987. *Neurofibromatosis* 1(3):172–178
133. Erlic Z, Rybicki L, Peczkowska M, Golcher H, Kann PH, Brauckhoff M, Mussig K, Muresan M, Schaffler A, Reisch N, Schott M, Fassnacht M, Opocher G, Klose S, Fottner C, Forrer F, Plockinger U, Petersenn S, Zabolotny D, Kollukch O, Yaremchuk S, Januszewicz A, Walz MK, Eng C, Neumann HP (2009) Clinical predictors and algorithm for the genetic diagnosis of pheochromocytoma patients. *Clin Cancer Res* 15(20):6378–6385
134. Neumann HP, Sullivan M, Winter A, Malinoc A, Hoffmann MM, Boedeker CC, Bertz H, Walz MK, Moeller LC, Schmid KW, Eng C (2011) Germline mutations of the TMEM127 gene in patients with paraganglioma of head and neck and extraadrenal abdominal sites. *J Clin Endocrinol Metab* 96(8):E1279–E1282
135. Yao L, Schiavi F, Cascon A, Qin Y, Inglada-Perez L, King EE, Toledo RA, Ercolino T, Rapizzi E, Ricketts CJ, Mori L, Giacche M, Mendola A, Taschin E, Boaretto F, Loli P, Iacobone M, Rossi GP, Biondi B, Lima-Junior JV, Kater CE, Bex M, Vikkula M, Grossman AB, Gruber SB, Barontini M, Persu A, Castellano M, Toledo SP, Maher ER, Mannelli M, Opocher G, Robledo M, Dahia PL (2010) Spectrum and prevalence of FP/TMEM127 gene mutations in pheochromocytomas and paragangliomas. *JAMA* 304(23):2611–2619
136. Neumann HP, Sullivan M, Winter A, Malinoc A, Hoffmann MM, Boedeker CC, Bertz H, Walz MK, Moeller LC, Schmid KW, Eng C (2011) Germline mutations of the TMEM127 gene in patients with paraganglioma of head and neck and extraadrenal abdominal sites. *J Clin Endocrinol Metab* 96(8):E1279–E1282. doi:[10.1210/jc.2011-0114](https://doi.org/10.1210/jc.2011-0114)
137. Burnichon N, Cascon A, Schiavi F, Morales NP, Comino-Mendez I, Abermil N, Inglada-Perez L, de Cubas AA, Amar L, Barontini M, de Quiros SB, Bertherat J, Bignon YJ, Blok MJ, Bobisse S, Borrego S, Castellano M, Chanson P, Chiara MD, Corssmit EP, Giacche M, de Krijger RR, Ercolino T, Girerd X, Gomez-Garcia EB, Gomez-Grana A, Guilhem I, Hes FJ, Honrado E, Korpershoek E, Lenders JW, Leton R, Mensenkamp AR, Merlo A, Mori L, Murat A, Pierre P, Plouin PF, Prodanov T, Quesada-Charneco M, Qin N, Rapizzi E, Raymond V, Reisch N, Roncador G, Ruiz-Ferrer M, Schillo F, Stegmann AP, Suarez C, Taschin E, Timmers HJ, Tops CM, Urioste M, Beuschlein F, Pacak K, Mannelli M, Dahia PL, Opocher G, Eisenhofer G, Gimenez-Roqueplo AP, Robledo M (2012) MAX mutations cause hereditary and sporadic pheochromocytoma and paraganglioma. *Clin Cancer Res* 18(10):2828–2837. doi:[10.1158/1078-0432.CCR-12-0160](https://doi.org/10.1158/1078-0432.CCR-12-0160)
138. Hong SB, Oh H, Valera VA, Stull J, Ngo DT, Baba M, Merino MJ, Linehan WM, Schmidt LS (2010) Tumor suppressor FLCN inhibits tumorigenesis of a FLCN-null renal cancer cell line and regulates expression of key molecules in TGF-beta signaling. *Mol Cancer* 9:160

139. Hudon V, Sabourin S, Dydensborg AB, Kottis V, Ghazi A, Paquet M, Crosby K, Pomerleau V, Uetani N, Pause A (2010) Renal tumour suppressor function of the Birt-Hogg-Dube syndrome gene product folliculin. *J Med Genet* 47(3):182–189
140. Toro JR, Wei MH, Glenn GM, Weinreich M, Toure O, Vocke C, Turner M, Choyke P, Merino MJ, Pinto PA, Steinberg SM, Schmidt LS, Linehan WM (2008) BHD mutations, clinical and molecular genetic investigations of Birt-Hogg-Dube syndrome: a new series of 50 families and a review of published reports. *J Med Genet* 45(6):321–331
141. Schmidt LS, Nickerson ML, Warren MB, Glenn GM, Toro JR, Merino MJ, Turner ML, Choyke PL, Sharma N, Peterson J, Morrison P, Maher ER, Walther MM, Zbar B, Linehan WM (2005) Germline BHD-mutation spectrum and phenotype analysis of a large cohort of families with Birt-Hogg-Dube syndrome. *Am J Hum Genet* 76(6):1023–1033
142. Pavlovich CP, Grubb RL 3rd, Hurley K, Glenn GM, Toro J, Schmidt LS, Torres-Cabala C, Merino MJ, Zbar B, Choyke P, Walther MM, Linehan WM (2005) Evaluation and management of renal tumors in the Birt-Hogg-Dube syndrome. *J Urol* 173(5):1482–1486
143. Menko FH, van Steensel MA, Giraud S, Friis-Hansen L, Richard S, Ungari S, Nordenskjold M, Hansen TV, Solly J, Maher ER (2009) Birt-Hogg-Dube syndrome: diagnosis and management. *Lancet Oncol* 10(12):1199–1206
144. Boris RS, Benhammou J, Merino M, Pinto PA, Linehan WM, Bratslavsky G (2011) The impact of germline BHD mutation on histological concordance and clinical treatment of patients with bilateral renal masses and known unilateral oncocyoma. *J Urol* 185(6):2050–2055
145. Rutten A, Burgdorf W, Hugel H, Kutzner H, Hosseiny-Malayeri HR, Friedl W, Propping P, Kruse R (1999) Cystic sebaceous tumors as marker lesions for the Muir-Torre syndrome: a histopathologic and molecular genetic study. *Am J Dermatopathol* 21(5):405–413
146. Lynch HT, de la Chapelle A (2003) Hereditary colorectal cancer. *N Engl J Med* 348(10):919–932
147. Umar A (2004) Lynch syndrome (HNPCC) and microsatellite instability. *Dis Markers* 20(4–5):179–180
148. Bocker T, Ruschoff J, Fishel R (1999) Molecular diagnostics of cancer predisposition: hereditary non-polyposis colorectal carcinoma and mismatch repair defects. *Biochim Biophys Acta* 1423(3):O1–O10
149. Entius MM, Keller JJ, Drillenburger P, Kuypers KC, Giardiello FM, Offerhaus GJ (2000) Microsatellite instability and expression of hMLH-1 and hMSH-2 in sebaceous gland carcinomas as markers for Muir-Torre syndrome. *Clin Cancer Res* 6(5):1784–1789
150. Machin P, Catusus L, Pons C, Munoz J, Conde-Zurita JM, Balmana J, Barnadas M, Marti RM, Prat J, Matias-Guiu X (2002) Microsatellite instability and immunostaining for MSH-2 and MLH-1 in cutaneous and internal tumors from patients with the Muir-Torre syndrome. *J Cutan Pathol* 29(7):415–420
151. Ponti G, Losi L, Di Gregorio C, Roncucci L, Pedroni M, Scarselli A, Benatti P, Seidenari S, Pellacani G, Lembo L, Rossi G, Marino M, Lucci-Cordisco E, Ponz de Leon M (2005) Identification of Muir-Torre syndrome among patients with sebaceous tumors and keratoacanthomas: role of clinical features, microsatellite instability, and immunohistochemistry. *Cancer* 103(5):1018–1025
152. Aarnio M, Sankila R, Pukkala E, Salovaara R, Aaltonen LA, de la Chapelle A, Peltomaki P, Mecklin JP, Jarvinen HJ (1999) Cancer risk in mutation carriers of DNA-mismatch-repair genes. *Int J Cancer* 81(2):214–218
153. Ponti G, Losi L, Pedroni M, Lucci-Cordisco E, Di Gregorio C, Pellacani G, Seidenari S (2006) Value of MLH1 and MSH2 mutations in the appearance of Muir-Torre syndrome phenotype in HNPCC patients presenting sebaceous gland tumors or keratoacanthomas. *J Invest Dermatol* 126(10):2302–2307
154. South CD, Hampel H, Comeras I, Westman JA, Frankel WL, de la Chapelle A (2008) The frequency of Muir-Torre syndrome among Lynch syndrome families. *J Natl Cancer Inst* 100(4):277–281
155. Schwartz RA, Torre DP (1995) The Muir-Torre syndrome: a 25-year retrospect. *J Am Acad Dermatol* 33(1):90–104
156. Chhibber V, Dresser K, Mahalingam M (2008) MSH-6: extending the reliability of immunohistochemistry as a screening tool in Muir-Torre syndrome. *Mod Pathol* 21(2):159–164
157. Kruse R, Rutten A, Schweiger N, Jakob E, Mathiak M, Propping P, Mangold E, Bisceglia M, Ruzicka T (2003) Frequency of microsatellite instability in unselected sebaceous gland neoplasias and hyperplasias. *J Invest Dermatol* 120(5):858–864
158. Cesinaro AM, Ubiali A, Sighinolfi P, Trentini GP, Gentili F, Facchetti F (2007) Mismatch repair proteins expression and microsatellite instability in skin lesions with sebaceous differentiation: a study in different clinical subgroups with and without extracutaneous cancer. *Am J Dermatopathol* 29(4):351–358
159. Orta L, Klimstra DS, Qin J, Mecca P, Tang LH, Busam KJ, Shia J (2009) Towards identification of hereditary DNA mismatch repair deficiency: sebaceous neoplasm warrants routine immunohistochemical screening regardless of patient's age or other clinical characteristics. *Am J Surg Pathol* 33(6):934–944
160. Jarvinen HJ, Aarnio M, Mustonen H, Aktan-Collan K, Aaltonen LA, Peltomaki P, De La Chapelle A, Mecklin JP (2000) Controlled 15-year trial on screening for colorectal cancer in families with hereditary nonpolyposis colorectal cancer. *Gastroenterology* 118(5):829–834
161. Kruse R, Ruzicka T (2004) DNA mismatch repair and the significance of a sebaceous skin tumor for visceral cancer prevention. *Trends Mol Med* 10(3):136–141
162. Lynch HT, Fusaro RM, Lynch PM (2006) Sebaceous skin lesions as clues to hereditary non-polyposis colorectal cancer. *J Invest Dermatol* 126(10):2158–2159
163. Schneider DT, Janig U, Calaminus G, Gobel U, Harms D (2003) Ovarian sex cord-stromal tumors—a clinicopathological study of 72 cases from the Kiel Pediatric Tumor Registry. *Virchows Arch* 443(4):549–560
164. Scully RE (1970) Sex cord tumor with annular tubules a distinctive ovarian tumor of the Peutz-Jeghers syndrome. *Cancer* 25(5):1107–1121
165. Jeghers H, Mc KV, Katz KH (1949) Generalized intestinal polyposis and melanin spots of the oral mucosa, lips and digits; a syndrome of diagnostic significance. *N Engl J Med* 241(26):1031–1036
166. Tomlinson IP, Houlston RS (1997) Peutz-Jeghers syndrome. *J Med Genet* 34(12):1007–1011
167. Giardiello FM, Welsh SB, Hamilton SR, Offerhaus GJ, Gittelsohn AM, Booker SV, Krush AJ, Yardley JH, Luk GD (1987) Increased risk of cancer in the Peutz-Jeghers syndrome. *N Engl J Med* 316(24):1511–1514
168. Giardiello FM, Brensinger JD, Tersmette AC, Goodman SN, Petersen GM, Booker SV, Cruz-Correa M, Offerhaus JA (2000) Very high risk of cancer in familial Peutz-Jeghers syndrome. *Gastroenterology* 119(6):1447–1453
169. Boardman LA, Thibodeau SN, Schaid DJ, Lindor NM, McDonnell SK, Burgart LJ, Ahlquist DA, Podratz KC, Pittelkow M, Hartmann LC (1998) Increased risk for cancer in patients with the Peutz-Jeghers syndrome. *Ann Intern Med* 128(11):896–899
170. Hearle N, Schumacher V, Menko FH, Olschwang S, Boardman LA, Gille JJ, Keller JJ, Westerman AM, Scott RJ, Lim W,

- Trimbath JD, Giardiello FM, Gruber SB, Offerhaus GJ, de Rooij FW, Wilson JH, Hansmann A, Moslein G, Royer-Pokora B, Vogel T, Phillips RK, Spigelman AD, Houlston RS (2006) Frequency and spectrum of cancers in the Peutz-Jeghers syndrome. *Clin Cancer Res* 12(10):3209–3215
171. Mehenni H, Resta N, Park JG, Miyaki M, Guanti G, Costanza MC (2006) Cancer risks in LKB1 germline mutation carriers. *Gut* 55(7):984–990
172. Lim W, Olschwang S, Keller JJ, Westerman AM, Menko FH, Boardman LA, Scott RJ, Trimbath J, Giardiello FM, Gruber SB, Gille JJ, Offerhaus GJ, de Rooij FW, Wilson JH, Spigelman AD, Phillips RK, Houlston RS (2004) Relative frequency and morphology of cancers in STK11 mutation carriers. *Gastroenterology* 126(7):1788–1794
173. Giardiello FM, Trimbath JD (2006) Peutz-Jeghers syndrome and management recommendations. *Clin Gastroenterol Hepatol* 4(4):408–415
174. van Lier MG, Wagner A, Mathus-Vliegen EM, Kuipers EJ, Steyerberg EW, van Leerdam ME High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations. *Am J Gastroenterol* 105(6):1258–1264; author reply 1265
175. Jenne DE, Reimann H, Nezu J, Friedel W, Loff S, Jeschke R, Muller O, Back W, Zimmer M (1998) Peutz-Jeghers syndrome is caused by mutations in a novel serine threonine kinase. *Nat Genet* 18(1):38–43
176. Hemminki A, Markie D, Tomlinson I, Avizienyte E, Roth S, Loukola A, Bignell G, Warren W, Aminoff M, Hoglund P, Jarvinen H, Kristo P, Pelin K, Ridanpaa M, Salovaara R, Toro T, Bodmer W, Olschwang S, Olsen AS, Stratton MR, de la Chapelle A, Aaltonen LA (1998) A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. *Nature* 391(6663):184–187
177. Aretz S, Stienen D, Uhlhaas S, Stolte M, Entius MM, Loff S, Back W, Kaufmann A, Keller KM, Blaas SH, Siebert R, Vogt S, Spranger S, Holinski-Feder E, Sunde L, Propping P, Friedl W (2007) High proportion of large genomic deletions and a genotype phenotype update in 80 unrelated families with juvenile polyposis syndrome. *J Med Genet* 44(11):702–709
178. Young RH, Welch WR, Dickersin GR, Scully RE (1982) Ovarian sex cord tumor with annular tubules: review of 74 cases including 27 with Peutz-Jeghers syndrome and four with adenoma malignum of the cervix. *Cancer* 50(7):1384–1402
179. (2003) American Society of Clinical Oncology policy statement update: genetic testing for cancer susceptibility. *J Clin Oncol* 21(12):2397–2406
180. (2011) American College of Surgeons: commission on cancer. Cancer program standards 2012: ensuring patient-centered care, vol V1.0